

**OSTEOPROTECTIVE EFFECT OF DIFFERENT COMPONENTS OF  
*MORINGA OLIEFERA* IN OVARIECTOMY INDUCED  
OSTEOPOROSIS MODEL OF WISTAR RATS**

**Pragna Parikh\*, Chirag Patel and Ayaz Rangrez**

Department of Zoology, Faculty of Science, The Maharaja Sayajirao University of Baroda,  
Vadodara.

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**\*Correspondence for  
Author**

**Prof. Pragna Parikh**

Department of Zoology,  
Faculty of Science, The  
Maharaja Sayajirao  
University of Baroda,  
Vadodara.

**ABSTRACT**

Osteoporosis is a bone condition defined by low bone mass, increased fragility, decreased bone quality, and an increased fracture risk. Due to lack of compliance in current pharmacological interventions targeting bone problems like postmenopausal osteoporosis, there is an urge for developing new alternative therapies for osteoporosis. In recent times, interest has been given to phytotherapy due their ease of availability and acquiescence. Hence, the present study demonstrates that supplementing ovariectomized (OVX) animals with *Moringa oliefera*

(MO) flower, leaf and fruit extract have positive effect on bone health. Following ovariectomy there was a dramatic decrease in the serum calcium levels, with an increase in the excretion of calcium. Elevated TRAcP and ALP after OVX decreased with MO extract exposure, fall in both the markers clearly suggest that MO plant extract ameliorate the damage caused by estrogen deficiency. Further, phytochemical analysis showed an array of phytochemicals with anti inflammatory as well as antioxidant properties, proving its osteoprotective efficacy. In general, the presence of these Phytochemicals could account for the much touted medicinal properties. Because of the chemical complexity of the MO, one individual phytochemical cannot be given the credit for its pharmacological property. Some compounds may be collectively affecting broad aspects of physiology, detoxification mechanisms, reducing the stress and re-supplementing the lost hormones such as phytoestrogens.

**KEYWORDS:** *Osteoporosis, Phytochemicals, Ovariectomy, Moringa Oliefera.*

## INTRODUCTION

Osteoporosis becomes a serious health threat for aging postmenopausal women by predisposing them to an increased risk of fracture. Osteoporotic fractures are associated with substantial morbidity and mortality in postmenopausal women, especially older women.<sup>[1-2-3]</sup> The pathogenesis of osteoporosis is associated with increased osteoclastic bone resorption which is due to the increase in the production of pro-inflammatory cytokines like IL-1 and IL-6.<sup>[5-6]</sup> These pro-inflammatory cytokines prevents the apoptosis of osteoclasts and diminishes the osteoblast proliferation. This may be attributed to sex steroid deficiency as well as increase in glucocorticoids. Identifying the pharmacological agents which can modulate the osteoclasts and osteoblast cells, so as to preserve or enhance bone mass is the main stay of current pharmacological treatments for osteoporosis. Various therapies are in use for the treatment of osteoporosis, including calcium supplementation with Vitamin D<sub>3</sub> and Hormone Replacement Therapy (HRT) with estrogen, selective estrogen receptor modulators, calcitonin, raloxifene, amino-bisphosphonates, teriparatide, parathyroid hormone, strontium ranelate, growth hormone, and IGF -1.<sup>[6]</sup> However, HRT poses the threat of breast, ovarian and endometrial cancer. Hence, alternative therapies are gaining attendance to discover new remedies for osteoporosis<sup>[6]</sup> One of such alternative therapy, phytotherapy, is earning the importance these days because of its lesser side effects and compliance. Various plants have the potency to check the activity of osteoclastic cells as well to promote osteoblastic cells. Medicinal plants constitute an effective source of traditional and modern medicines.<sup>[7]</sup> Phytotherapy is gaining its importance in the new world countries and is considered to be more effective, safe and compliant when compared to classical drugs like strontium renelate or even HRT. *Litsea glutinosa*, an ayurvedic herb, has been proved to ameliorates calcium metabolism, increases the osteoblastic number as well as activity and prevents the osteoclastic bone resorption in overactomised rats.<sup>[8-9]</sup> It has been reported by Bureau of plant industry that *Moringa* is an outstanding source nutritional components. Asserting its multi-faceted value, the plant is utilized for its highly nutritive, medicinal, and water purification properties.<sup>[10]</sup> The plant's nutritive properties are ubiquitous throughout the plant, resulting in the observation that most plant parts attain nutrition and can be eaten: leaves, seeds, bark, roots, exudates, flowers, and pods.<sup>[11-12]</sup> A list of possible medical applications conferred by *M. oleifera* plant parts includes, antihypertensive, anticancer, antispasmodic, antitumor, antiulcer, cholesterol lowering, diuretic, hepatoprotective, and hypoglycemic capabilities, as well as treatment of infectious skin and mucosal diseases.<sup>[13]</sup> Leaf extracts have been used to treat hyperthyroidism and currently fruits of *moringa* have

been proved to have osteoprotective effect on ovariectomized Wistar rats<sup>[14]</sup> however the studies were confined to leaves only. Hence The objectives of the present study was to evaluate the effect of ethanolic extract of different parts of *moringa oliefera* (Fruit, Flower and Leaf) and to analyze the phyto components of all the three parts of MO for its usefulness in preventing bone loss in estrogen deficient OVX rats.

## MATERIALS AND METHODS

### Experimental protocol

The experimental protocol was approved by IAEC (Institutional animal ethical committee). 3 month-old virgin female Wistar rats 30 in number were procured from Sun Pharma Advance Research Center (300±20gm). The animals were acclimatized for 8 days before the onset of the experiment to adapt to laboratory conditions (The room temperature was 22±4 °C with a 12 h/12 h light/dark cycle). Then the rats were ovariectomized (OVX) and sham operated after being anesthetized under intraperitoneal injection of sodium pentobarbital at a dose of 30mg/kg body weight, as described previously.<sup>[15]</sup> The success of the OVX was assessed through vaginal cytology after five days of surgery as described previously<sup>[16]</sup>. Rats were given the lag phase of 10 days to recover from the stress of operation and then treatment was started as follows.

Group 1: Sham control and received vehicle.

Group 2: Ovariectomized (OVX) control and received vehicle.

Group 3: Ovariectomized + leaf extract (200 mg/kg b.wt. / day oral)

Group 4: OVX + flower extract (200 mg/kg b.wt./day / oral)

Group 5: OVX + fruit extract (200 mg/kg b.wt. /day /oral). (LD<sub>50</sub>=400mg/kg B.W).

### Preparation of Extract and phytochemical analysis

Fruits, leaves and flowers of MO were obtained following the method of.<sup>[9]</sup> All the parts of the plants were dry in oven at 50° C until constant weight was attained. They were kept away from direct sunlight to avoid destroying active compounds. They were then minced using automated mincer and fine powder was produced. Dried powder was prepared by drying MO in oven at 50° C. 100 gm dried powder of each component was extracted with 500 ml methanol in Soxhlet's apparatus for 48 hours . Methanolic extract was dried on water bath at 55° C. The percentage yield of the plant was found to be 9.8%, 6.3% and 7.7% for fruits, leaves and flowers respectively. The plant extract was freeze dried and stored at -70° C. Working solution was prepared by dissolving the extract in DMEM and filtered using 0.23 μ

filters. The dry extracts of MO leaves, flower and fruit were used to determine the compounds. The qualitative methods for flavanoids, alkanoids steroids and triterpenoids, saponins, Tanins and Phenols was performed using standered method.<sup>[17-22]</sup>

### **Gas chromatographic analysis**

GC/MS analysis was carried out using Perkin Elmer autosystem XL with turbo mass system equipped with PE 5 MS 30m X 250 micron silica capillary. Injector and detector temperatures were 250° and 300°C, respectively. The temperature started from 70° C for 5 min and then rose to 290° C at the rate of 10° C per minute. Helium was used as carrier gas. The MS was taken at 70 eV. Scanning speed was 0.84 scans s<sup>-1</sup> and the scanning period was from 40 to 550 s. Sample volume was kept 3 µL.

### **Biochemical analysis**

Every alternate day food and water intake was measured, and the body weight was recorded. All experimental designs and procedures had received the approval of the institutional ethics committee. The treatment was continued for 30 days and at the end of experimental period total urine excreted over 24 h period was collected from overnight fasted rats by housing each group individually in a metabolic cage. Animals were euthanized by overdose of ether; blood was collected by orbital sinus puncture. 0.5 ml blood was collected; serum was separated and stored at -80° C for further analysis. Rats were dissected, bone, uterus and liver was immediately removed, washed in PBS (pH 7.4), and stored at -80° C for further analysis. Liver and uterus were blotted and weighed. All the assays were carried out using commercial kits purchased from Reckon Diagnostics.

### **Statistical Analysis**

Data were expressed as mean values and S.E.M. One-way ANOVA was used to compare data from all groups and Bonferroni post test to compare the results ( $p < 0.05$  by the statistical software of Graph Pad PRISM (Version 5.0)). A  $p$  value of less than 0.05 was considered statistically significant.

## **RESULTS**

### **Body weight and relative organ weight**

As shown in Table 1, after 8 weeks of study there was significant increase in the body weight of all the OVX animals, whereas control animals showed a marginal increase in the weight. Exposure of MO extract to OVX resulted into marginal decrease in the body weight, and of

the three parts of MO extracts used, exhibited significant decrease in the body weight (Figure 1). Maximum alterations were observed in relative uterine weight. As expected, ovariectomy caused significant decrease in the weight of the uterus and relative uterine weight (Table 2). However, no changes were observed in the relative uterine weight of any of the treatment groups (Figure 2).

### **Serum and urine biochemical Markers**

Table III represents the alterations in the serum and urine biochemical parameters. OVX resulted into a decrease in serum calcium levels 50% (Table III). Decrease in the calcium was poorly shielded by leaf extracts of MO; however, flower and fruit extracts improved the serum calcium profile (Figure 5). OVX resulted into an enhanced excretion of calcium in the urine, reaching almost 3 fold calcium compared to control (Table III), and treatment with MO components reduced the loss of calcium in the serum, (Figure 8). OVX led to increased urinary excretion which was enhanced to 3 fold at the end of 2 months. MO Leaf extract did not show any significant effect on the calcium excretion, MO flower and fruit extract treatment reduced the calcium excretory rate, (Figure 8). As far as phosphorus is concerned an increase in phosphate level was reported in OVX rats, which seen to be improved by all the three components of MO extract (Figure 6). Serum ALP, the functional markers of osteoblast activity was observed to increase in OVX, MO Plant extracts was successful in improving serum ALP levels (Figure 3). Serum TRAcP, an osteoclast specific marker was found to be high in all OVX rats, and MO extract treatment resulted into lowering the activity (Figure 4).

### **Tissue biochemical Markers**

Bone AIP levels were significantly higher in the OVX group compared to normal and MO plant extract treatment showed lowered AIP levels with all the three components. However, maximum effect was observed with fruit and flower extracts of MO (Figure 9 and 10). Bone TRAcP as anticipated, were higher in OVX animals compared to normal. All the three components of MO plant extract reduced the TRAcP activity in bone .However, liver ALP (Figure 11) didn't show any significant alterations, thus the ALP is Serum can be correlated with that of Bone.

### **Phytochemical analysis**

The phytochemical screening of plants gave positive results for flavonoids, alkaloids, phenols, anthocyanins and saponins. Both flower and fruit was found to be rich in flavonoids,

phenols and saponins compared to leaves. Preliminary tests also confirmed that leaves are having presence of tannins, but we were not able to detect the presence of tannins in flower and fruits (Table IV). GC MS profile of leaf, flower and fruit extract were studied. Figure 18, 19 and 20 shows the GC scan of leaf, flower and fruit. Relative abundance was measured and the phytochemicals were detected by their comparison of mass spectra with standard mass library. The list of phytochemicals detected using GC MS are listed in Table V.

**Table I: Increase in body weight in different treatment groups**

	Control	Ovx	Leaf	Flower	Fruit
1	242.500± 5.280	248.330±3.330	240.10± 4.170	240.000±3.330	240.000±3.330
2	250.000± 6.110	260.830± 3.610	245.23±3.890	245.830±3.890	247.830± 3.890
3	262.500±3.330	270.830±5.830	262.43± 3.610	266.23±3.330	252.500±3.330
4	266.670± 3.610	285.830± 4.170	266.15± 3.890	275.000±3.060	268.000± 3.060
5	273.330± 4.170	295.830±5.000	275.45± 4.170	284.170±2.500	276.170±2.500
6	282.500± 3.890	305± 4.170	282.32 ± 5.280	291.830±5.000	283.670± 5.000
7	291.670± 5.000	315 ± 2.780	291.07± 3.610	295.000±2.220	298± 4.170
8	295.830± 5.280	332.500±5.280	305.30± 5.830	307.10±4.170	310 ± 2.780

Values were expressed as Mean ± S.E.M. \* - p < 0.05; \*\* - p < 0.01; \*\*\* - p < 0.001.

**Table II: Relative uterine weight in different experimental groups.**

	Control	Ovx	OVX+Leaf	OVX+Flower	OVX+Fruit
Relative uterine weight	0.001± 0.0001	0.00022**±0.000152	0.00024±0.000225	0.000235±0.000205	0.000235±0.000205

Values were expressed as Mean ± S.E.M. \* - p < 0.05; \*\* - p < 0.01; \*\*\* - p < 0.001.

**Table III: Serum/Urine Biochemical markers**

Parameters	Control	Ovx	Ovx +Leaf	Ovx +Flower	Ovx +Fruit	
Serum Profile	AIP	54.663± 4.57	124.569***± 6.33	118.455± 6.336	79.258***±8.336	68.456***±4.336
	Calcium	8.811± 0.189	5.465***± 0.262	6.608± 0.229*	8.466***±0.293	7.456**±0.233
	Phosphate	7.910±0.569	18.223***±1.892	10.336***±0.689	11.569***±0.881	10.236***±0.233
	TRAcP	6.583±0.556	15.166***± 0.651	9.336*±0.812	7.986**± 0.556	8.456*±1.265
Urinary calcium	4.325± 0.986	12.669***±1.336	11.566±1.256	8.569*± 2.336	7.255* ±1.236	
Calcium Excretion rate	0.518±0.110	1.307**± 0.235	0.977± 0.145	0.618*± 0.105	0.655*± 0.129	

Values were expressed as Mean ± S.E.M. \* - p < 0.05; \*\* - p < 0.01; \*\*\* - p < 0.001.

**Table IV: Bone and Liver biochemical markers**

Treatment groups	control	Ovx	Ovx +Leaf	Ovx +Flower	Ovx +Fruit	
Bone	AIP	140.5±5.1	168.6**±4.2	155.6*±8.3	146.6**±7.3	144.6**±6.3
	TRAcP	6.583± 0.556	15.166***±0.651	9.336*± 2.669	7.986**± 0.669	8.456**± 1.365
Liver	AIP	79.081± 12.367	82.1±20.593	79.335±18.844	77.176± 19.883	78.127± 24.734

Values were expressed as Mean ± S.E.M. \* - p < 0.05; \*\* - p < 0.01; \*\*\* - p < 0.001

Table V: Phytochemical analysis of MO

Fraction	Test	Leaf	Flower	Fruit	
Flavonoids	N-lead acetate	+	+++	+++	
	Zinc dust	+	+++	+++	
	NaOH	++	+++	+++	
	H <sub>2</sub> SO <sub>4</sub>	+	+++	+++	
Terpenoids	Chl+H <sub>2</sub> so <sub>4</sub>	+	+	+	
Phenols	N-FeCl <sub>3</sub>	+	++	+++	
	FeSO <sub>4</sub>	++	+++	+++	
Alkaloids	Mayers	++	+	+	
	Wagners	-	+	+	
	Dragondorff's	++	+	+	
Anthocyanins	Na acetate	-	+	+	
	Na <sub>2</sub> CO <sub>3</sub>	-	+	+	
Tannins	Gelatin	+	-	-	
	K <sub>2</sub> Cr <sub>2</sub> O <sub>7</sub>	+	-	-	
	Iodine	+	-	-	
	Lead acetate	+	-	-	
Saponin	Water	+	+++	+++	
	Lead acetate	+	+++	+++	
Amines	Na-Nitro	-	-	-	
	Dragondorff's	-	-	-	
	Ehlich	-	-	-	
Glycosides	cynogenic	Molisch	+	-	+
		Cold H <sub>2</sub> SO <sub>4</sub>	-	+	+
	cardiotonic	Kedde	-	-	-
		Keller	-	+	+

Key: + Low concentration; ++ Moderate concentration; +++ High concentration; - absent (negative)

Table VI : Phytochemicals detected using GC MS scan in different components of MO

LEAF	FLOWER	FRUIT
Adenosine derivatives	Pyridine carbonitrile	Stearic acid derivatives
Ergolin	Thiazolamine	Decanoic acid derivatives
Teretrahydroquinoline	Tertra hydroquinoline	Piperizine derviations
Cinnamic acid	Phenanthridone	Olean derivatives
Oleic acid derivatives	Cinnoline	Glucobrassicin
Piperizininie derivatives	Cinnamic acid	Octadecane
Heneicosane	Coumarin	Palmitic acid derivatives
Eicosane	Ribitol	Quinolizine derivatives
Quebrachamine	Olien derivatives	Triprolidine
Curan	Ergolin	Ibogamine
Thebaine	Androstan	Yohimbane
Aspidospermidine	Ibogamine	
Carnegine	Nonahexacontanoic acid	
	Arabinitol	
	Aristolocholic acid	

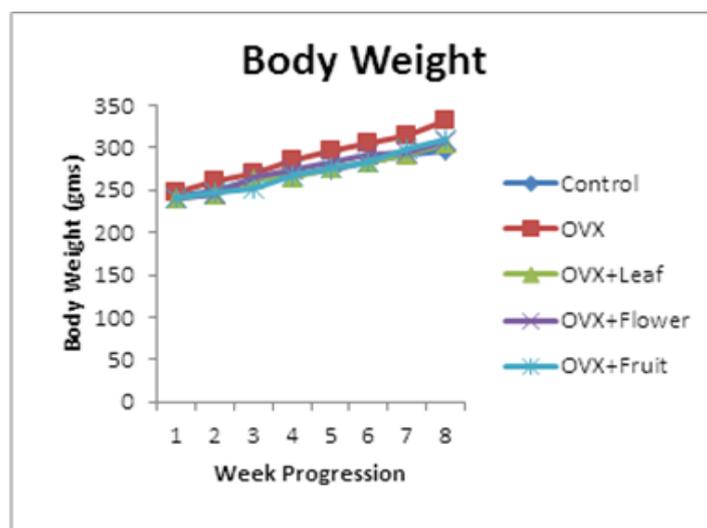


Figure 1: Comparison of Body weight of Rats treated with MO components with Control

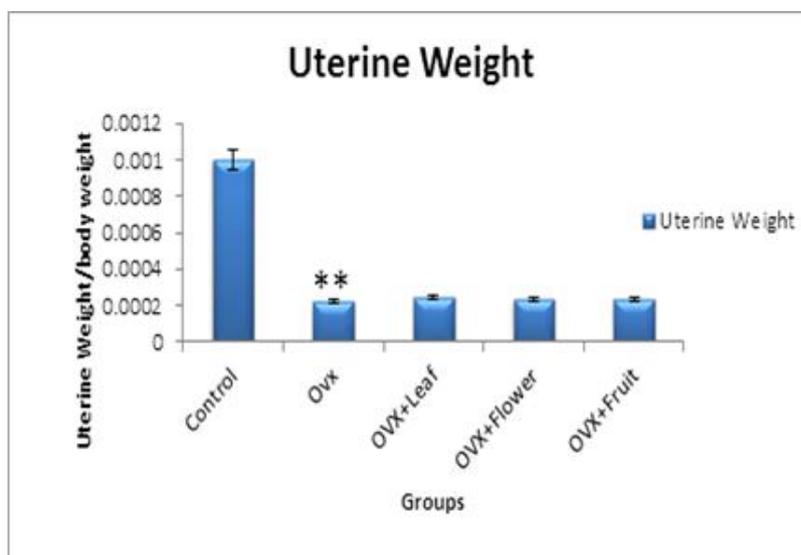


Figure 2: Comparison of Uterine weight of Rats treated with MO components with Control

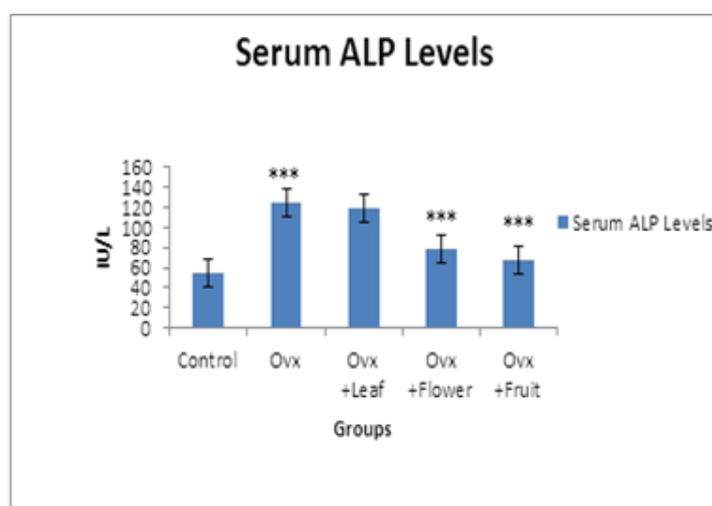


Figure 3: Alteration in Serum ALP Levels in different groups treated MO extracts

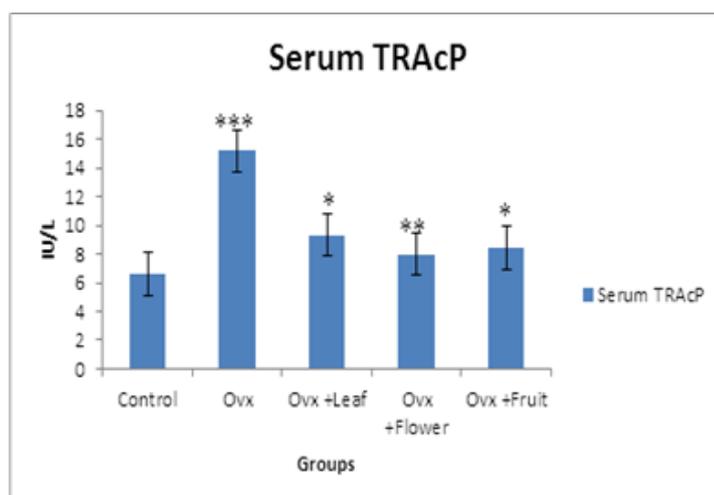


Figure 4: Alteraion in Serum TRAcP Levels in different groups treated MO extracts.

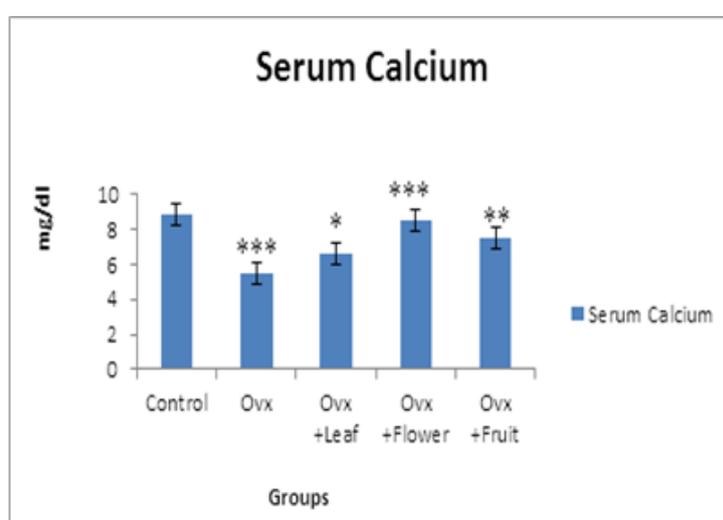


Figure 5: Alteraion in Serum Calcium Levels in different groups treated MO extracts.

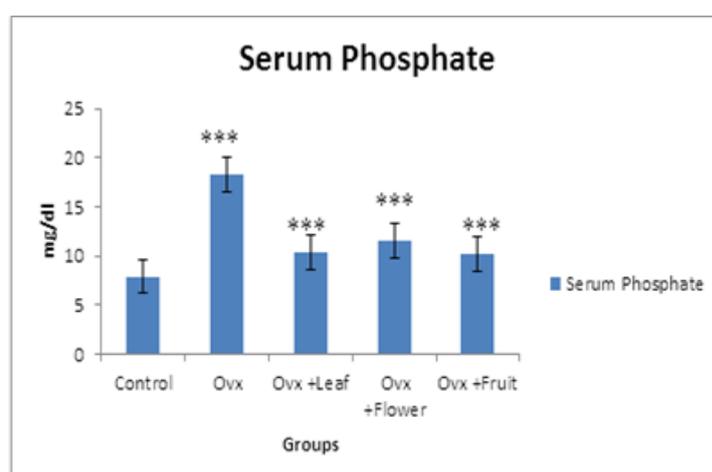


Figure 6: Alteraion in Serum Phosphate Levels in different groups treated MO extracts.

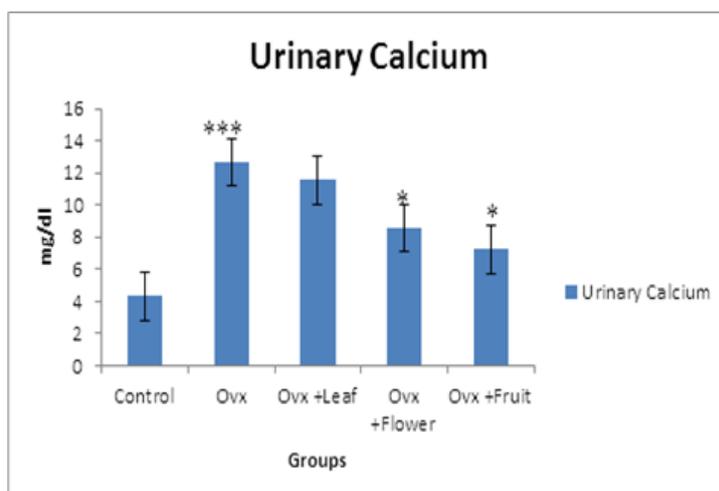


Figure 7: Alteraion in Urinary Calcium Levels in different groups treated MO extracts.

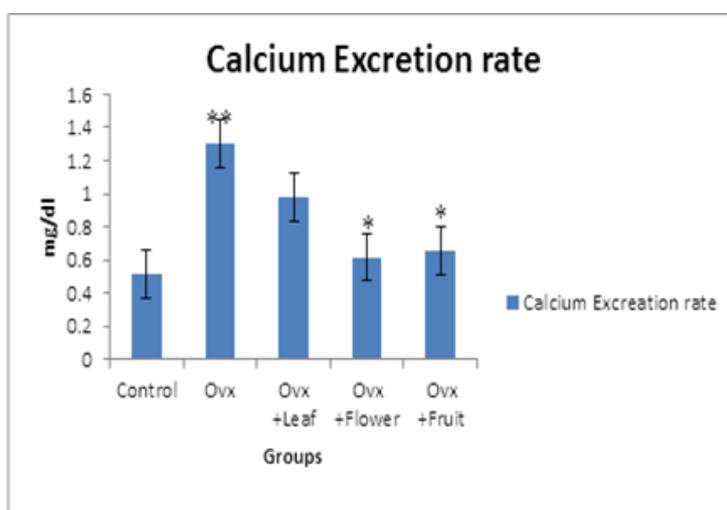


Figure 8: Alteraion in Serum Phosphate Levels in different groups treated MO extracts.

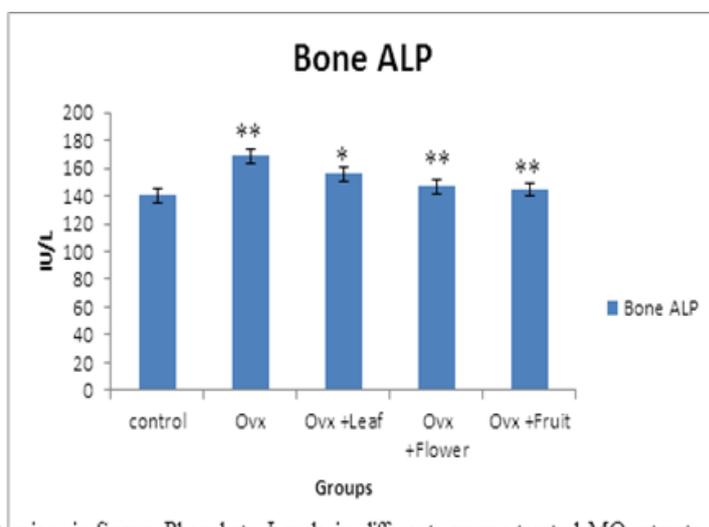


Figure 9: Alteraion in Serum Phosphate Levels in different groups treated MO extracts.

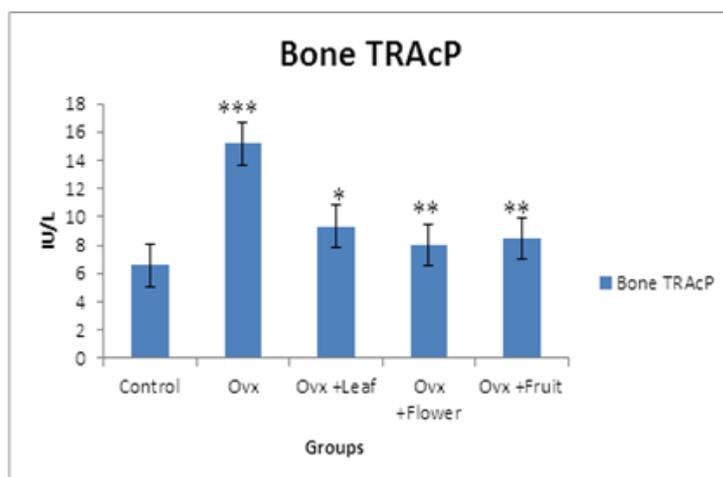


Figure 10: Alteration in Bone TRAcP Levels in different groups treated MO extracts.

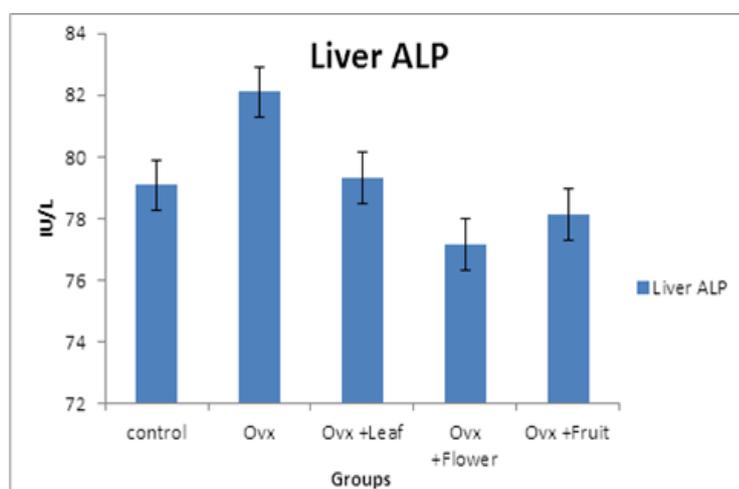


Figure 11: Alteration in Liver ALP Levels in different groups treated MO extracts.

## DISCUSSION

The present study demonstrated that supplementing OVX animals with MO flower, leaf and fruit extracts can have positive effect on bone health. OVX animals showed weight gain which may be due to lack of estrogen. MO flower and fruit had non-significant effect in reducing the weight gain. Maximum weight reduction was observed with leaf extract treatment, which may be due to presence of tannins in leaf of MO. Tannins have been reported to be responsible for decreases in feed intake, growth rate, feed efficiency, net metabolizable energy, and protein digestibility in experimental animals<sup>[23]</sup> tannins present in the extract could potentially inhibit the activity of lipases found in rats, thereby lowering their fat content.<sup>[24-26]</sup> The results indicate that the rats treated with MO leaf extract have significantly attenuated the body weight proving the hypolipidemic and thermogenic property.<sup>[27-28]</sup>

Ovariectomy induced absence of sex hormone initially affects the uterus, leading to uterine atrophy and decrease in the uterine weight, similar results was observed, where the uterine weight was decreased in OVX animals. MO treatment had no significant effects on the uterine. Our observations are in accordance to with the work done by<sup>[29-30]</sup> who have opined that plants ameliorates OVX induced changes on the bone without affecting the uterus. In the present study the MO extract also did not have the effect on the uterus but potentially inhibits the progression of post menopausal symptoms.

Decrease in serum calcium and increase in urinary calcium post ovariectomy is an established fact and one of the possible reasons for excess bone resorption.<sup>[31]</sup> Our study also showed similar results and further when excretory rate was considered, it was observed that calcium excretory rate was very high in OVX animals. Ovariectomy in the rat leads to an increase in intestinal calcium secretion, leading to impaired calcium balance, probably this high excretory rate affects the overall calcium metabolism and negative calcium metabolism leads to further progression of the disease<sup>[32]</sup> Of the three components of MO, leaf was having significant effect on improving the fall in serum calcium Similar observation were seen in case of flower and fruit extracts which were having more significant effects on boosting serum calcium compare to leaf extracts. When we considered the calcium excretory rate, it was clear that MO plant extract, had significant effects on calcium excretion, where, urine calcium excretion as well as the calcium excretory rated were seen to be decreasing suggesting that the components of the extracts were helping in keeping up the calcium load required by the OVX rats. Furthermore, MO plant extracts in OVX rats did not only restored the decreased serum calcium levels but also the phosphorus levels were seen to be near to normal, suggesting that MO flower and fruit extracts are effective in inhibiting bone resorption and in increasing bone formation. Numerous studies have shown the health-promoting properties of polyphenols, through which they promote skeletal health by reducing resorption caused by high oxidative stress.<sup>[33-35]</sup> The presence of polyphenols in MO plant thus suggests its role is reducing resorption. The antioxidant properties of polyphenols have also been widely studied and reported in the literature.<sup>[36-38]</sup> They strongly support the role of polyphenols in the delayed onset or reduction in the progression of osteoporosis. The protective effects of polyphenols against diseases, including osteoporosis, have generated new expectations for improvements in health. Our findings are also in agreement with the findings<sup>[39-41]</sup> who reported that plants containing polyunsaturated fatty acids increases serum

calcium and phosphorus concentrations and reduce urinary calcium and phosphorous excretion, thus enhanced bone formation.

Previous studies in our lab have established that bone is maintained through coordinated activities of osteoblasts and osteoclasts and ovariectomy disrupts this balance, leading to over-activity of osteoclasts, followed by osteoblasts.<sup>[8]</sup> After a lag phase, osteoblasts are not able to keep up with the pace of osteoclasts, leading to an imbalance, where the bone is resorbed much more than it can be synthesized. Two functional markers of this process are AIP and TRAcP, which are secreted by osteoblasts and osteoclasts respectively.<sup>[42,9]</sup> In the present study we observed that following ovariectomy, there was an increase in serum AIP and TRAcP. MO extract exposure led to a fall in ALP as well as TRAcP levels, possibly due to the presence of flavonoids as proposed by<sup>[43]</sup> that certain flavonoids can have positive effect on bone nodule formation. All the three parts of MO plant was found to be rich in flavonoids which might be the one playing a key role in stimulating osteoblast/osteoclast cells. Of the three components maximum effects was seen with the flower, proving its beneficial role in preventing further progression of osteoporosis in OVX rats. Parallel fall in both the markers clearly suggest that fruit, flower and leaf extracts have improved the damage caused by estrogen deficiency. To confirm that the elevation in serum AIP levels is because of bone remodeling only, we estimated liver AIP levels also, which showed no significant variation between the MO extract treated group and control group.

Thus the present study has established the osteoprotective efficacy of leaf, fruit and flower of MO plant. The phytochemical profile of MO leaf, flower and fruits are different and they exhibit different pharmacological property. Studies have proved that<sup>[44]</sup> and cinnamic acid<sup>[45]</sup> are osteoprotective. As MO components are rich in cinnamic acid, probably its osteoprotective efficacy is enhanced by the presence of it. Presence of PUFA and flavonoids also participate in expression of the beneficial effect on bone health.<sup>[46]</sup> Apart from this, another botanical agent Coumarin which is found in flower of MO is also attributed to promote bone health (Tang *et al.*, 2008). The Proliferation and maturation of Osteoclasts follows a pathway that involves variety of inflammatory cytokines. As all the parts of the MO plant shows the presence of flavonoids and hence, the robust anti inflammatory activity might be directly affecting the inflammatory pathways of development of Osteoclasts.<sup>[47-48]</sup>

## CONCLUSION

Our Study is the first to prove the effect of MO Components on OVX induced Rats and showed that among the entire component tested, Flower and fruit gave most promising results. Further, the elucidation of phytochemical compositions from MO extracts, it validated the presence of osteoprotective compounds, which upon isolation can be use as herbal therapeutics for this type of bone disease.

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## REFERENCES

1. The North American Menopause Society. The management of osteoporosis in postmenopausal women, position statement of The North American Menopause Society. *Menopause* 2006; 13: 340-367.
2. NAMS continuing medical education activity Management of osteoporosis in postmenopausal women: *Menopause: The Journal of The North American Menopause Society*, 2006; 17(1): 24-59.
3. Nozaka K, Miyakoshi N, Kasukawa Y, Maekawa S, Noguchi H, Shimada Y. Intermittent administration of human parathyroid hormone enhances bone formation and union at the site of cancellous bone osteotomy in normal and ovariectomized rats. *Bone*, 2008; 42(1): 90–97.
4. Xie F, Wu CF, Lai WP, Yang XJ, Cheung PY, Yao XS, Leung PC, Wong MS.. The osteoprotective effect of Herba epimedii (HEP) extract *in vivo* and *in vitro*. *Evid. Based Complement. Alternat. Med*, 2005; 2: 353-361.
5. Park JA, Ha SK, Kang TH, Oh MS, Lee M H, Lee SY. Protective effect of apigenin on ovariectomy induced bone loss in rats. *Life Sciences*, 2008; 82: 1217-1223.
6. Xie MH, Aggarwal S, Ho WH, Foster J, Zhang Z, Stinson J, Wood WI, Goddard AD, and Gurney, A.L. Interleukin (IL)-22, a novel human cytokine that signals through the interferon receptor-related proteins CRF2-4 and IL-22R. *J. Biol. Chem.*, 2000; 275: 31335–31339.
7. Mahmood A, Ahmad M, Jabeen A, Zafar M, Nadeem S. Pharmacognostic Studies of Some Indigenous Medicinal Plants of Pakistan. *Ethnobotanical Leaflets.*, 2005; 9: 1.

8. Parikh P, Suresh B, Rangrez A. Osteoprotective effect of *Litsea glutinosa* in ovariectomized wistar rats. *Electronic Journal of Pharmacology & Therapy*, 2009; 2: 81-86.
9. Rangrez A, Suresh B, Parikh PH. Osteoprotective effect three anti inflammatory plants in ovariectomized wistar rats *Pharmacologyonline*, 2011; 1: 675-684.
10. Anwar F, Sajid L, Muhammad A, Anwarul HG. 2007. *Moringa oleifera*: A Food plant with Multiple Medicinal Uses. *Phytother. Res*, 2007; 21: 17-25.
11. Fahey JW. *Moringa oleifera*: A Review of the Medical Evidence for Its Nutritional Therapeutic and Prophylactic Properties. Part 1. *Trees for Life Journal.*, 2005; 1: 5.
12. Kamal T, Muzammil A, Abdullateef RA, Muhammad NO. Investigation of antioxidant activity and phytochemical constituents of *Artocarpus altilis*, *Journal of Medicinal Plants Research*, 2012; 6(26): 4354-4357,
13. Rockwood JL, Anderson BG, Casamatta DA. Potential uses of *moringa oleifera* and an examination of antibiotic efficacy conferred by *m. oleifera* seed and leaf extracts using crude extraction techniques available to underserved indigenous populations. *International Journal of Phytotherapy Research*, 2013; 3(2): 61-71.
14. Burali SC, Kangralkar V, Sravani O, Patil SL. The beneficial effect of ethanolic extract of *moringa oleifera* on osteoporosis. *International journal of pharmaceutical applications*, 2010; 1(1): 50-58.
15. Xiao XL, HARA I, Matsumiya T. Effects of osthole on postmenopausal osteoporosis using ovariectomized rats; comparison to the effects of Estradiol. *Biol. Pharm. Bull*, 2002; 25(6): 738—742.
16. Roveri EA, Chap G, Grappiolo I, Puche RC. Effects of depot medroxyprogesterone acetate on the calcium metabolism of adult ovariectomized rats. *Medicine*, 2000; 60: 482-486.
17. Ciulei. *Practical Manuals on the Industrial Utilization of Medicinal And Aromatic Plants*, Romania, University of Bucharest. *Clinical Endocrinology and Metabolism*, 1964; 86: 5217-5221.
18. Chitravadivu C, Manian S, Kalachelvi K. Qualitative analysis of Selected Medicinal Plants, Tamilnadu, India. *Middle East J. Sci. Res*, 2009; 4: 144-146.
19. Devmurari VP, Ghodasara TJ, Jivani NP. Antibacterial Activity and Phytochemical Study of Ethanolic Extract of *Triumfetta rhomboidea* Jacq. *International Journal of PharmTech Research*, 2010; 2(2): 1182-1186.

20. Kam TS, Sim KM, Lim TM. Voasstrictine, a novel pentacyclic quinolinic alkaloid from *Tabernaemontana*. *Tetrahedron Lett*, 2001; 42: 4721-4723.
21. Daniel M. Method. In: *Plant Chemistry and Economic Botany*. Kalyani publishers New Delhi., 1991; 63-64.
22. Sahu VK, Irchhaiya R, Shashi A, Gurjar H. Phytochemical investigation and chromatographic evaluation of the ethanolic extract of whole plant extract of *Dendrophthoe falcata* (L.f.) ettingsh. *Int. Jr. Pharma Sci and Research*, 2010; 1(1): 39-45.
23. Muthusamy K, Gopalakrishnan S, Ravi Tk, Sivachidambaram P. Biosurfactants: properties, commercial production and application. *Current Science*, 2008; 94: 736-746.
24. Christine L. Chichioco-Hernandez, Finella Marie G. Leonido. Weight-lowering effects of *Caesalpinia pulcherrima*, *Cassia fistula* and *Senna alata* leaf extracts. *Journal of Medicinal Plants Research.*, 2011; 5(3): 452-455,
25. Khan S, MirzaKJ, Abdin MZ. Development of RAPD markers for authentication of medicinal plant *Cuscuta reflexa*, *EurAsia J Bio Sci.*, 2010; 4: 1-7.
26. Ikechukwu CE, Obri A, Igiri A. The effect of aqueous ethanolic extract of *Stachytarpheta cayennensis* on the histology of the liver and fasting blood sugar of non-diabetic and diabetic wistar rats. *The international journal of nutrition and wellness*, 10(1)
27. Waterman A S, Schwartz SJ, Zamboanga BL, Ravert R D, Williams MK, Agocha V B. The Questionnaire for Eudaimonic Well-Being: Psychometric properties, demographic comparisons, and evidence of validity. *J. Posit. Psychol.*, 2010; 5: 41–61
28. Bais S, Singh GS, Sharma R. “Antiobesity and Hypolipidemic Activity of *Moringa oleifera* Leaves against High Fat Diet-Induced Obesity in Rats.” *Advances in Biology*, 2014; 9.
29. Roveri EA, Chap G, Grappiolo I, Puche RC. Effects of depot medroxyprogesterone acetate on the calcium metabolism of adult ovariectomized rats. *Medicine*, 2000; 60: 482-486.
30. Xie F, Wu CF, Lai WP, Yang XJ, Cheung PY, Yao XS, Leung PC, Wong MS. The osteoprotective effect of *Herba epimedii* (HEP) extract *in vivo* and *in vitro*. *Evid. Based Complement. Alternat. Med*, 2005; 2: 353-361.
31. Zhang M, Xuan S, Boussein ML, von Stechow D, Akeno N, Faugere MC, Malluche H, Zhao G, Rosen CJ, Efstratiadis A, Clemens TL. Osteoblast-specific knockout of the insulin-like growth factor (IGF) receptor gene reveals an essential role of IGF signaling in bone matrix mineralization. *J Biol Chem*, 2002; 277: 44005-44012.

32. Akao M, Abe R, Sato N, Tanigome AH, Kumagai H, Hitomi Kumagai. Prevention of Osteoporosis by Oral Administration of Phytate-Removed and Deamidated Soybean  $\beta$ -Conglycinin. *Int. J. Mol. Sci*, 2014; 16: 2117-2129.
33. Trzeciakiewicz A. When nutrition interacts with osteoblast function: molecular mechanisms of polyphenols. *Nutrition Research Reviews*, 2009; 22: 68-81.
34. Tucker KL. Osteoporosis Prevention and Nutrition. *Current Osteoporosis Reports*, 2009; 7(4): 111.
35. Hunter DC, Skinner M A, Lister CE. Impact of phytochemicals on maintaining bone and joint health. *Nutrition (Burbank, Los Angeles County, Calif)*, 2008; 24(4): 390-392.
36. Rassi CM, Lieberherr M, Chaumaz G, Pointillart and courtot G. Down regulation of osteoclast differentiated by diadzein via caspase 3. *j. bon min res*. 2002; 17: 630
37. Shen CL, Cao JJ, Dagda RY, Tenner TE, Jr MC, Yeh J K. Supplementation with green tea polyphenols improves bone microstructure and quality in aged, orchidectomized rats. *Calcified Tissue International*, 2011; 88(6): 455-463.
38. Rao AV, Rao LG. Carotenoids and human health. *Pharmacological Research*, 2007; 55(3): 207-216.
39. Qin L. Gene expression profiles and transcription factors involved in parathyroid hormone signaling in osteoblasts revealed by microarray and bioinformatics. *J. Biol. Chem*, 2003; 278: 19723–19731.
40. Reddy K, Kulkarni KS. The Efficacy of OST-6, A Polyherbal Formulation in the Management of Primary Osteoporosis : A Pilot Study, *Orthopaedics Today (IV)*, 2002; 3: 195-199.
41. Zhang Y, Lai WP, Leung PC, Wu CF, Yao XS, Wong MS. Effects of Fructus Ligustri Lucidi extract on bone turnover and calcium balance in ovariectomized rats. *Biol. Pharm. Bull*, 2006; 29: 291-296.
42. Burmeister B, Domaschke H. Co culture of osteoclast and osteoblast on resorbable mineralized collagen scaffolds: Establishment of an in vitro model of bone remodeling. *European cells and materias*, 2003; 5(2): 18-19.
43. Vali B, Rao LG, El-Sohehy. Epigallocatechin-3-gallate increases the formation of mineralized bone nodules by human osteoblast-like cells. *Journal of Nutritional Biochemistry*, 2007; 18(5): 341–347.
44. Tang C, Yang R, Chin M. Enhancement of bone morphogenic protein 2 expression and bone formation by Cumarine deteirvative via p38 and ERk dependant pathway in osteoblasts. *European Journal of Phamachology*, 2008; 576: 40-49.

45. Lai KA, Shen WJ, Yang CY, Shao CJ, Hsu JT, Lin RM. The use of alendronate to prevent early collapse of the femoral head in patients with non traumatic osteonecrosis: A randomized clinical study. *J Bone Joint Surg Am*, 2005; 87: 2155–9.
46. Penolazzi L, Vecchiati R, Bignard S, Lambertini E, Torreggiani E, Canella A, Franceschetti T, Calura G, Vesce F, Piva R. Influence of obstetric factors on osteogenic potential of umbilical cord-derived mesenchymal stem cells. *Reproductive Biology and Endocrinology*, 2009; 7: 106
47. Owusu-Ansah M, Achel DG, Adaboro RM, Asarel DK, Amoatey HM. Total Phenolic Content and Antioxidant Activity in Leaf Samples of Twelve Accessions of *Moringa oleifera* Lam. *Journal of Chemical and Analytical Science*, 2011; 2(10): 1226-1230.
48. Luqman S, Srivastava S, Kumar R, Maurya AK, Chanda D. Experimental Assessment of *Moringa oleifera* Leaf and Fruit for Its Antistress, Antioxidant, and Scavenging Potential Using *In Vitro* and *In Vivo* Assays. *Evidence-Based Complementary and Alternative Medicine.*, 2012; 12.