



**A COMPARATIVE STUDY ON ANTI DIABETIC ACTIVITY OF CRUDE EXTRACTS OF  
ENDOPHYTIC FUNGI FROM SEA WEED GRACILARIA CORTICATA**

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**ABSTRACT**

Diabetes, often referred to by doctors as Diabetes mellitus, describes a group of metabolic diseases in which the person has high blood glucose (blood sugar). Insulin is a hormone that is produced by the pancreas. Inhibition of  $\alpha$ -amylase, enzyme that plays a role in digestion of starch and glycogen, is considered a strategy for the treatment of disorders in carbohydrate uptake, such as diabetes. Gracilaria corticata have been used since ancient times as food, fodder, and fertiliser and as source of medicine. Gracilaria corticata. Endophytic fungi are one of the potential natural resources for new ant diabetic compound sources. The aim of the study was to compare anti diabetic activity of crude extracts of Endophytic fungi from Gracilaria corticata against diabetes. Endophytic fungi were isolated from seaweed Gracilaria corticata, characterized and identified based on morphological and biochemical characters. The identified endophytic fungi were tested for invitro antidiabetic activity by alpha amylase inhibition assay, method the highest percentage of alpha amylase inhibition at the concentration of 500 $\mu$ l of ethanolic extract of A. niger that showed 41.4% followed by penicillium (39%) A.fumigatus (33.4%) Phoma spp (28.9%) and Syncephalastrum racemosum (23.7%) in comparison with acetone crude extracts of Penicillium (25.2%), Phoma spp (24.2%), A.fumigatus (21.3%), Syncephalastrum racemosum (19.4%), A.niger (17.4%).

**KEYWORDS:** Endo phytic fungi, DNS, Gracilaria corticata, Diabetes, Alpha amylase.

**INTRODUCTION**

Diabetes (Diabetes mellitus) is classed as a metabolism disorder. Metabolism refers to the way our bodies use digested food for energy and growth. Most of what we eat is broken down into glucose. Glucose is a form of sugar in the blood - it is the principal source of fuel for our bodies. The body does not produce enough insulin for proper function, or the cells in the body do not react to insulin (insulin resistance). Approximately 90% of all cases of diabetes worldwide are of this type. Some people may be able to control their type 2 diabetes symptoms by losing weight, following a healthy diet, doing plenty of exercise, and monitoring their blood glucose levels. However, type 2 diabetes is typically a progressive disease - it gradually gets worse - and the patient will probably end up have to take insulin, usually in tablet form. Overweight and obese people have a much higher risk of developing type 2 diabetes compared to those with a healthy body weight. People with a lot of visceral fat, also known as central obesity, belly fat, or abdominal obesity, are especially at risk. Gestational diabetes affects females during pregnancy. Some women have very high levels of glucose in their blood, and their bodies are unable to produce enough insulin to transport

all of the glucose into their cells, resulting in progressively rising levels of glucose. Diagnosis of gestational diabetes is made during pregnancy

**ROLE OF ALPHA AMYLASE IN DIABETES**

Amylase plays an important role in digestion of starch in to glucose in Blood of Human body. When starch is consumed amylase is secreted and converts in to Sugar. Insulin is the hormone produced by Pancreas to inhibit alpha amylase. In certain cases insulin is not enough to inhibit alpha amylase activity hence leads to increase in blood glucose levels. The main aim of the study was to inhibit alpha amylase by using crude extracts of Endophytic fungi isolated from Gracilaria corticata.

**SEA WEED – GRACILARIA CORTICATA**

Gracilaria corticata is a sea weed which belongs and haposses well branched thalli and are multicellular. They survive in marine environments on rocks and appears fleshy. Gracilaria species accomodate medically and commercially important metabolites.

### ENDOPHYTIC FUNGI

Endophytic fungi are the excellent resources for metabolites against Diabetes. Endophytic fungi are organisms found in plants, sea weeds etc. They grow within the tissues producing secondary metabolites exhibiting antidiabetic activities without affecting the host cells.

### MATERIALS AND METHODS

The *Gracilaria corticata*, a sea Weed was collected from Kovalam Beach, Chennai and processed by washing with tap water to remove salts and other adhering particles. The washed sea weeds were immersed in 80% ethanol for 3 mins and rinsed with sterile distilled water three times for 10 seconds and allowed to surface dry under sterile conditions. After drying, the sea weeds were cut into segments approximately 0.5 cm squares and placed on petri plates.

### ISOLATION OF ENDOPHYTIC FUNGI

The cut sea weeds were placed on to PDA and SDA plates incubated at 30°C for 2 weeks. They were monitored everyday for the growth of endophytic fungal colonies. Fungi growing out from the samples were subsequently transferred to fresh PDA plates. The procedure of transferring to fresh PDA plates and finally to slant for several times in order to isolate pure colonies.

### FERMENTATION

The endophytic fungus was grown on potato dextrose yeast agar (PDYEA) at 30°C for 5-7 days depending on growth rate. Loopful of grown culture from the PDA slant were inoculated into 1000 ml Erlenmeyer flasks containing 500 ml potato dextrose yeast extract broth (PDYEB) and incubated at 30°C for 4 weeks. After incubation period, the fungal cultures were harvested and filtered through two layers of cheese cloth. The dried mycelium was extracted three times with ethanol and acetone. The solvent was evaporated to dryness under reduced pressure to obtain a crude extract.



### ANTI DIABETIC ACTIVITY BY ALPHA AMYLASE INHIBITION ASSAY METHOD

250 µl of α amylase solution (1mg/ml phosphate buffer) was added to 100 µl of sample using blank solution and mixed well. The tubes were incubated at 37°C for 20 mins in water bath. 250 µl of substrate solution (0.5%

starch in phosphate buffer) was added and mixed well followed by incubation at 37°C for 15 mins. 2ml of DNS (Dinitrosalicylic acid reagent): (40 mM DNS, sodium potassium tartrate, 0.4% M NaOH) was added to stop the reaction. Mix well and boil at 100°C for 10 mins. Cool the mixture and measure the absorbance in spectrophotometer at 540 nm.

### RESULTS

The morphological characteristics of the fungal isolates were observed and described according to their standard taxonomic key included



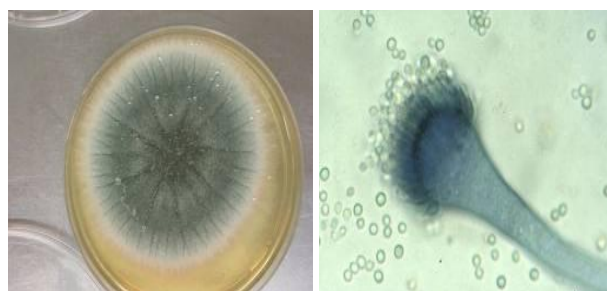
Macroscopic and Microscopic view of *A. niger*.



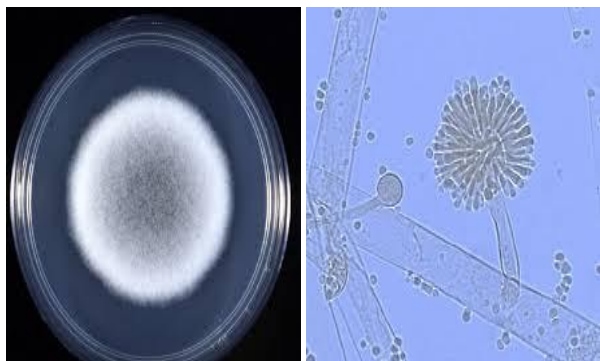
Macroscopic and microscopic view of *Penicillium* sps.



Macroscopic and microscopic view of *Phoma* sps.



Macroscopic and microscopic view of *A. fumigatus*.



Macroscopic and microscopic view of *Syncephalastrum racemosum*.

ACETONE EXTRACT

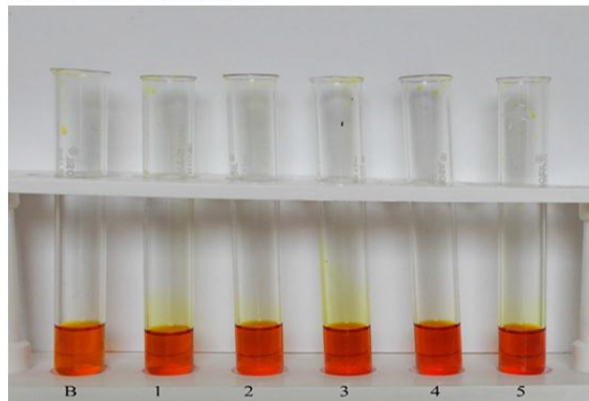
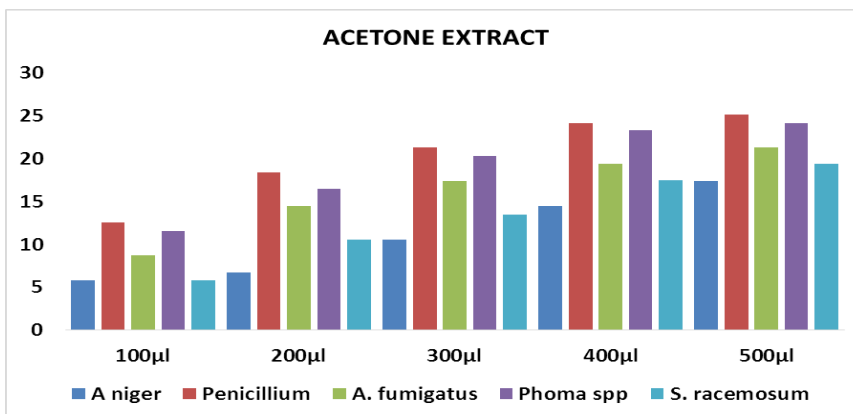


FIGURE: 1.

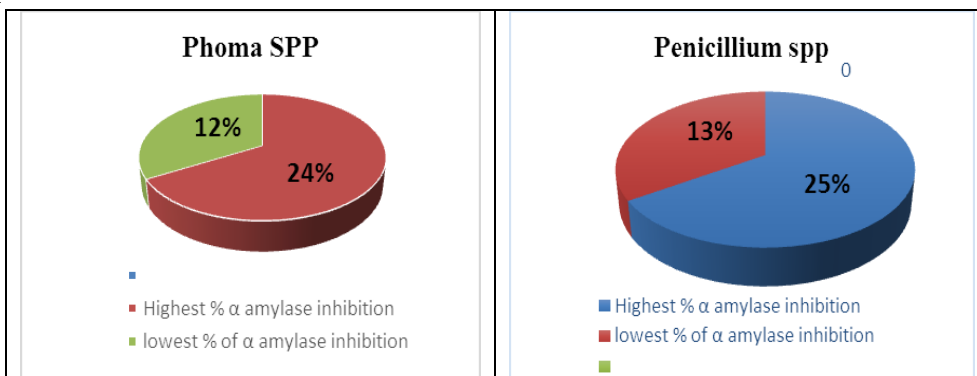
TABLE: 1.

| ENDOPHYTIC FUNGI                 | % of $\alpha$ amylase Inhibition |             |             |             |             |
|----------------------------------|----------------------------------|-------------|-------------|-------------|-------------|
|                                  | 100 $\mu$ l                      | 200 $\mu$ l | 300 $\mu$ l | 400 $\mu$ l | 500 $\mu$ l |
| <i>Aspergillus niger</i>         | 5.8                              | 6.7         | 10.6        | 14.5        | 17.4        |
| <i>Penicillium spp</i>           | <b>12.6</b>                      | 18.4        | 21.3        | 24.2        | <b>25.2</b> |
| <i>Aspergillus fumigatus</i>     | 8.7                              | 14.5        | 17.4        | 19.4        | 21.3        |
| <i>Phoma spp</i>                 | 11.6                             | 16.5        | 20.3        | 23.3        | 24.2        |
| <i>Syncephalastrum racemosum</i> | 5.8                              | 10.6        | 13.5        | 17.4        | 19.4        |



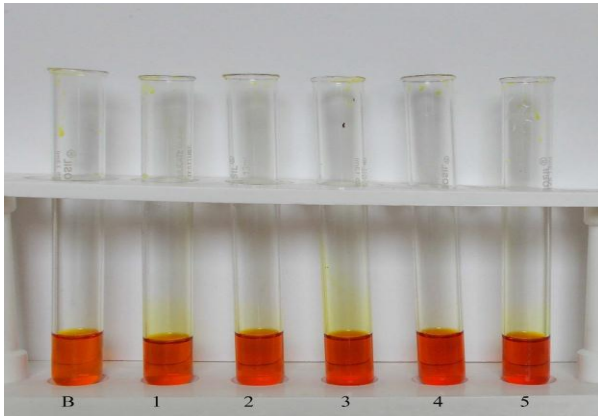
| FUNGI                  | HIGHEST % OF $\alpha$ AMYLASEINHIBITION | LOWEST % OF $\alpha$ AMYLASEINHIBITION |
|------------------------|---|--|
| <i>Penicillium spp</i> | 25%                                     | 13%                                    |
| <i>Phoma spp</i>       | 24%                                     | 12%                                    |

PIE CHART



The above table and chart showed the highest and lowest percentage of inhibition of alpha amylase by crude acetone extract of *Penicillium* and *Phoma* spp.

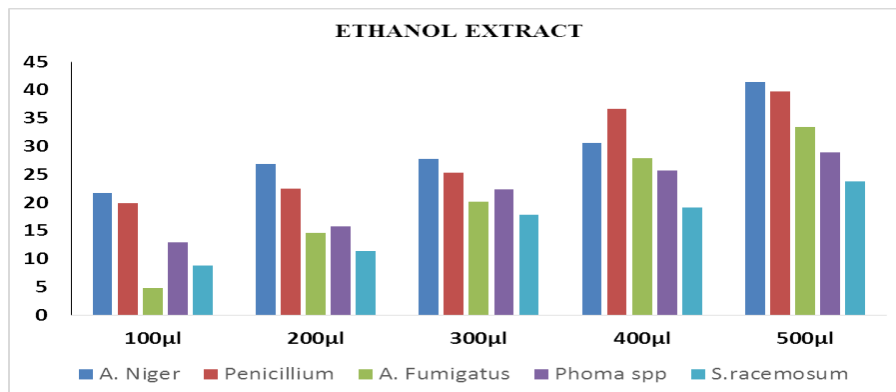
**ETHANOL EXTRACT**



**FIGURE: 2.**

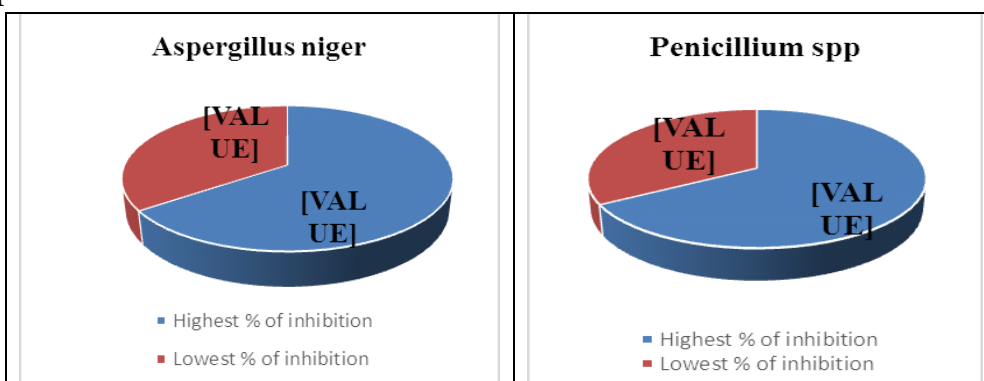
**TABLE: 2.**

| ENDOPHYTIC FUNGI          | % of $\alpha$ amylase Inhibition |             |             |             |             |
|---------------------------|----------------------------------|-------------|-------------|-------------|-------------|
|                           | 100 $\mu$ l                      | 200 $\mu$ l | 300 $\mu$ l | 400 $\mu$ l | 500 $\mu$ l |
| Aspergillus niger         | 21.7                             | 26.8        | 27.8        | 30.6        | 41.4        |
| Penicillium spp           | 19.9                             | 22.5        | 25.3        | 36.7        | 39.8        |
| Aspergillus fumigatus     | 4.8                              | 14.6        | 20.2        | 27.9        | 33.4        |
| Phoma spp                 | 12.9                             | 15.8        | 16.3        | 17.7        | 19.1        |
| Syncephalastrum racemosum | 8.8                              | 11.4        | 17.8        | 19.1        | 23.7        |



| FUNGI             | HIGHEST % OF $\alpha$ AMYLASEINHIBITION | LOWEST % OF $\alpha$ AMYLASEINHIBITION |
|-------------------|---|--|
| Aspergillus niger | 41%                                     | 22%                                    |
| Penicillium       | 40%                                     | 20%                                    |

**PIE CHART**

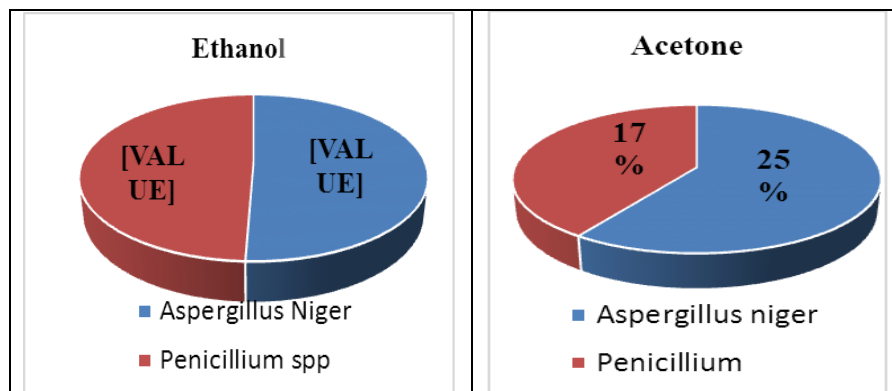


The above table and chart showed the highest and lowest percentage of inhibition of alpha amylase by crude ethanolic extract of Aspergillus niger and Penicillium.

### COMPARISON OF ALPHA AMYLASE INHIBITORY ACTIVITY BY CRUDE ETHANOL AND ACETONE EXTRACTS OF ENDOPHYTIC FUNGI

| FUNGI             | ETHANOL EXTRACT | ACETONE EXTRACT |
|-------------------|-----------------|-----------------|
| Aspergillus niger | 41%             | 17%             |
| Penicillium       | 40%             | 25%             |

#### PIE CHART



#### DISCUSSION

The present work resulted in exhibiting the highest percentage of alpha amylase inhibition at the concentration of 500 $\mu$ l of ethanolic extract of *A. niger* that showed 41.4% followed by *penicillium* (39%) *A.fumigatus* (33.4%) *Phoma spp* (28.9%) and *Syncephalastrum racemosum* (23.7%) in comparison with acetone crude extracts of *Penicillium* (25.2%), *Phoma spp* (24.2%), *A.fumigatus* (21.3%), *Syncephalastrum racemosum* (19.4%), *A.niger* (17.4%).

#### CONCLUSION

Based on alpha amylase inhibition assay by different endophytic fungi isolated from sea weed *Gracilaria corticata* it was found that crude ethanolic extract of *Aspergillus niger* showed highest percentage of inhibition followed by *Penicillium spp*, the crude acetone extracts of *Penicillium* showed highest percentage followed by *Phoma spp*. The crude ethanolic extract of *Aspergillus niger* showed maximum percentage of alpha amylase inhibition compared to crude acetone extract of *penicillium Sps*.

#### ACKNOWLEDGEMENT

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#### REFERENCES

- Danaei G, Finucane MM, Lu Y, Singh GM, Cowan MJ, Paciorek CJ, *et al.* National, regional, and global trends in fasting plasma glucose and diabetes prevalence since 1980: systematic analysis of health examination surveys and epidemiological studies with 370 country-years and 2.7 million participants. *Lancet*, 2011; 378: 31–40.
- Zhen Sun, Feng Chen. Evaluation of the green algae *Chlorella pyrenoidosa* for the management of diabetes. *J Food Drug Anal*, 2012; 20: 246-9.
- International Diabetes Federation. IDF diabetes atlas. 6<sup>th</sup> edition. [www.idf.org/diabetesatlas](http://www.idf.org/diabetesatlas), 2013.
- Anjana RM, Pradeepa R, Deepa M, Datta M, Sudha V, Unnikrishan R, *et al.* For the ICMR-INDIAB collaborative Study Group. Prevalence of diabetes and pre-diabetes in urban and rural India: phase I results of ICMR-INDIAB STUDY. *Diabetol*, 2011; 54: 3022-7.
- American Diabetes association. Standards of medical care for patients with diabetes mellitus (position statement). *Diabetes Care*, 2002; 25: S33.
- Kathleen Mahan, Sylvia Escott-Stmp. Krause's food, Nutrition and diet therapy, Elsevier, USA, 2004.
- Shailimavardhini RD, Reddinaik B, Neelima M, Ramesh B. Screening and production of  $\alpha$ -amylase from *Aspergillus niger* using zero value material for solid state fermentation. *Int J Pharm Pharm Sci.*, 2005; 5: 55-60.
- Sindhu S, Nair, Vaibhavi Kavrekar, Anshu Mishra. *In vitro* studies on alpha amylase and alpha glucosidase inhibitory activity of selected plant extracts. *Eur J Exp Biol.*, 2013; 3: 128-32.
- Bhat M, Zinjarde SS, Bhargava SY, Kumar AR, Joshi BN. Antidiabetic Indian plants: a good source of potent amylase Inhibitors. *J Evidence-Based Complementary Altern Med.*, 2011; 810. <http://dx.doi.org/10.1093/ecam/nen040>
- Tarling CA, Woods K, Zhang R, Brastanos HC, Brayer GD, Andersen RJ, *et al.* The search for novel human pancreatic alpha-amylase inhibitors: high-through screening of terrestrial and marine natural product extract. *Chem Biol.*, 2008; 9: 433-8.
- Hasan Z, Yam MF, Ahemad Yusof AP. Anti-diabetic properties and mechanism of action of *Gynura procumbens* water extract in

- streptozotocin-induced diabetic rats. *Molecules*, 2010; 15: 9008-23.
12. Liu L, Yu YL, Liu C, Wang XT, Liu XD, Xie L. Insulin deficiency induces abnormal increases in intestinal disaccharides activities and expression under diabetic states: evidences from *in vivo* and *in vitro* study. *Biochem Pharmacol.*, 2011; 82: 1963-70.
  13. Sun HH, Mao WJ, Jiao JY, Xu JC, Li HY, Chen Y, *et al.* Structural characterization of extracellular polysaccharides produced by the marine fungus *Epicoccun nigrum* JJY-40 and their antioxidant activities. *Mar Biol.*, 2011; 13: 1048-55.
  14. Kamaladhasan N, Subramanian SK. Influence of seaweed liquid fertilizers on legume crop, red gram. *J Basic Appl Biol.*, 2009; 3: 21-4.
  15. Selvaraj K, Murugalakshmikumari R, Ramasubramanian V. Bio removal of nickel using seaweed as bio adsorbent. *J Basic Appl Biol.*, 2010; 4: 207-12.
  16. Domettila C, Brintha TSS, Sukumaran S, Jeeva S. Diversity and distribution of seaweeds in the muttom coastal waters, south-west coast of India. *Biodivers J.*, 2013; 4: 105-1.