



IN VIVO SCREENING OF ANALGESIC AND ANTIULCER ACTIVITY ON CARUM CARVI SEEDS

V. Swathi^{*1}, V. Sathish Kumar², Dr. Sk. Abdul Rahaman³ and Dr. Anjana Male⁴

¹Department of Pharmacology, Nirmala College of Pharmacy, Mangalagiri, Guntur, India.

²Pharm.D, Nirmala College of Pharmacy, Mangalagiri, Guntur, India.

^{3,4}Department of Chemistry, Nirmala College of Pharmacy, Mangalagiri, Guntur, India.

*Corresponding Author: V. Swathi

Department of Pharmacology, Nirmala College of Pharmacy, Mangalagiri, Guntur, India.

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ABSTRACT

The present study was designed to investigate the antiulcer and analgesic potential of *carum carvi* seeds. Caraway seeds use is common in diet, so there is no need for special administration of the drug and by little increase in intake quantity, the use of analgesics is abolished. Antiulcer activity was also evaluated by aspirin induced ulcer models. Effect of concurrent administration of ethanolic and aqueous extracts of seeds extract *Carum carvi* at a dose of 100 and 200 mg/kg b.w. respectively was given by oral route. Ethanolic and aqueous extracts of *Carum carvi* seeds significantly reduction in gastric content, total acidity, ulcer index, and increase in pH of gastric pylorus ligation ulcer model. In comparison with the standard drug, the results of hydro alcoholic extract at 100 mg dose showed good analgesic & at 200 mg dose showed antiulcer activity compared with a standard drug. Extracts of *Carum carvi* may be useful as a natural analgesic in the treatment of ulcer, inflammation, and pain.

KEYWORDS: *Carum carvi*, antiulcer activity, aspirin, and analgesic.

INTRODUCTION

An analgesic or painkiller is any member of the group of drugs used to achieve analgesia, relief from pain. A number of herbal drugs are used to treat Analgesia. Several epidemiological studies from different countries have reported widely varying prevalence rates for chronic pain, ranging from 12 to 80% of the population.^[6] It becomes more common as people approach death. A study of 4,703 patients found that 26% had pain in the last two years of life, increasing to 46% in the last month.^[2] The intensity of chronic pain was higher for girls, and girls' reports of chronic pain increased markedly between ages 12 and 14.^[3] Perception gives information on the pain's location, intensity, and something about its nature. The various conscious and unconscious responses to both sensation and perception, including the emotional response, add further definition to the overall concept of pain.^[4] Pain perception also varies depending on the location of the pain. The kinds of stimuli that cause a pain response on the skin include pricking, cutting, crushing, burning, and freezing. These same stimuli would not generate much of a response in the intestine. Intestinal pain arises from stimuli such as swelling, inflammation, and distension.^[4] Pain is the main reason for visiting the emergency department in more than 50% of cases and is present in 30% of family practice visits.^[5] Several epidemiological studies from different countries have reported widely

varying prevalence rates for chronic pain, ranging from 12 to 80% of the population.^[6] It becomes more common as people approach death. A study of 4,703 patients found that 26% had pain in the last two years of life, increasing to 46% in the last month.^[7] The intensity of chronic pain was higher for girls, and girls' reports of chronic pain increased markedly between ages 12 and 14.^[8] The success of NSAIDs in treating various inflammatory conditions such as rheumatoid arthritis (RA) and osteoarthritis (OA) validated inhibition of the enzyme prostaglandin H synthase (PGHS) or cyclooxygenase (COX) as a highly suitable target in anti-inflammatory therapies^[9,10] The following NSAIDs are derived from fenamic acid. Which is a derivative of anthranilic acid^[11] which in turn is a nitrogen isostere of salicylic acid, which is the active metabolite aspirin^[11] The first purified preparation of COX enzyme was reported in 1976^[12] More than a decade later COX was cloned in 1988^[13,14,15] In the 1990's and inducible isoform now called COX-2 was discovered.^[16,17,18] Inflammation (Latin, inflammation) is part of the complex biological response of body tissues to harmful stimuli, such as pathogens, damaged cells, or irritants.^[32]

Caraway (*Carum carvi*), also known as meridian fennel,^[19,20,21,22] or Persian cumin,^[31,33] is a biennial plant in the family Apiaceae,^[23] native to western Asia, Europe, and northern Africa. it occurs in India to a

limited extent in the temperate regions of the western Himalayas, Jammu and Kashmir and in chakratha hills of Uttar Pradesh. However, polyploid variants (with four haploid sets=4n) of this plant that was found to be perennial. Finland supplies approximately 28% (2011) of the world's caraway production.^[24] Caraway seeds are a rich source of dietary fiber. 100 g seeds provide 38 g of fiber, 100% of daily recommended intake of fiber. The fruits, usually used whole, have a pungent, anise-like flavor and aroma that comes from essential oils, mostly carvone, limonene,^[25] and anethole.^[26] It is used in caraway seed cake, and it is frequently added to sauerkraut.^[27,28,29,30] The roots may be cooked as a vegetable like parsnips or carrots. Additionally, the leaves are sometimes consumed as herbs, either raw, dried, or cooked, similar to parsley.^[31]

MATERIALS AND METHODS

Extraction and Preparation of Test Sample: The seeds of caraway were collected, dried and grinded into fine powder. 250 grams of caraway powder was weighed and extracted by using two types of solvents. Extraction was carried out by using 500ml of aqueous and hydroalcohol respectively by using maceration process. Until the complete extraction takes place. The extract obtained was filtered by using vacuum filtration and further, concentrated by using evaporation. The extract was further dried under air.

Albino Wistar strain rats obtained from Mahaveera Enterprises, Peerzadiguda, Ghatkesar Mandal, R. R. Dist-39. Weighing 150-250 gm of either sex were used in this study. Animals were acclimatized to laboratory conditions before testing e.g., room temperature of 25 ± 1°C, relative humidity 45-55% and a 12:12 h light dark cycle.

The experimental protocol was approved by the Institutional Animal Ethics Committee (IAEC) and protocol approval No. is 011/IAEC/Ncpa/Researchproposal/2015/16.

The Committee for Purpose of Control and Supervision of Experiments on Animals (CPCSEA) Reg. No. is 1629/po/a/12/CPCSEA. All the experimental procedures were carried out according to the proper guidelines of CPCSEA.

Analgesic Studies

Tail Immersion Method: In present study analgesia was assessed according to the method of Luiz *et al.* The animals fasted for 12 hours before the procedure was started. A total of six rats was grouped into four groups. One group treated with distilled water serves as control. The second group was treated Diclofenac sodium which acts as a standard. Other two groups were given test drugs i. e., caraway extract of 100mg/kg and 200 mg/kg. The drug is administered through oral route by using oral feeding needle. They were held in position in a suitable restrainer with a tail extending out. 3-4 cms of

the tail were marked and immersed in water bath thermostatically maintained at 55°C. The withdrawal time of tail from hot water (in seconds) was noted as reaction time or tail flick latency. The maximum cutoff time for immersion was 15 seconds to avoid the injuries of tissues of the tail. The initial reading was taken immediately before administration of the test and standard drugs and then 15, 30, 45, 60 and 75 minutes after the administration.

Anti-Ulcer Screening Method

Antiulcer activity was evaluated by an aspirin-induced ulcer in Wistar rats.^[8]

Aspirin-induced gastric ulcer model:

The animals fasted for 24 hours before the experiment. Animals were divided into 6 groups each containing 6 animals. Group 1 served as a positive control and received omeprazole at the dose of 30 mg/kg body weight, group 2 served as a negative control received aspirin at the dose of 500 mg/kg body weight. Group 3 received the aqueous extract of caraway seeds at the dose of 100 mg/kg body weight, Group 4 received the aqueous extract of caraway seeds at the dose of 200 mg/kg body weight, Group 5 received the hydroalcoholic extract of caraway seeds at the dose of 100 mg/kg body weight and Group 6 received the hydroalcoholic extract of caraway seeds at the dose of 200 mg/kg body weight. After one hour of extract administration, aspirin at a dose of 500 mg/kg body weight was administered orally to all four groups. Then after 6 hours animals were euthanized with an excess of chloroform and the stomach was excised and cut along the greater curvature, washed and the inner surface was examined for ulceration. Ulcer index and % ulcer protection were calculated by using the methods described in table-5,6. The ulcer index was then calculated by adding the total number of ulcers per stomach and total severity of ulcers per stomach.

Ulcer Index

Procedure: Animals in the group of aspirin induced ulcer were starved for 24 hours. The pylorus is ligated and the abdominal wall is closed with sutures. 3 hours after pyloric ligation, the animals were sacrificed. The abdomen was opened and a ligature was placed around the esophagus. The stomach was removed and fixed on a cork plate and the number of and severity of ulcers was registered with a stereomicroscope using the following scores

Ulcer index was calculated using following formula:

$$\text{Ulcer Index} = \frac{\text{(Arithmetic mean of (number of ulcers the intensity in a group) positive animals)}}{\text{Total number of animals}}$$

Determination of Total Acidity and Free Acidity Procedure

0.1mL of gastric juice specimen was transferred in a porcelain evaporating dish. 1-2 drops of Topfer's reagent is added. A colour change was observed; a bright red colour appears if free hydrochloric acid is present. 1-2

drops of phenolphthalein were added to the gastric juice with Topfer's reagent. Titrated with 0.01N NaOH from a burette, mixing was done after each addition until the last trace of red colour disappeared and was replaced by a canary yellow colour. The numbers of milliliters of NaOH used was read from the burette. This represents the amount of free hydrochloric acid. The titration was continued until the red colour of phenolphthalein appeared (deep pink), titrated to the point at which the further addition of alkali did not deepen the colour. Reading was taken (mL NaOH) for total acidity.

Calculation

$$Y = \text{mL of } 0.1 \text{ N NaOH} \times 10$$

Where, Y = Total acidity (mEq/L)

Acidity = Volume of NaOH x Normality x 100 mEq/litre

Acid volume

Procedure

Under ether anesthesia, a midline incision is made. Herbal extract was given to rats as per dosages and after a span of 1-hour aspirin was given at a dose of 500mg/kg body weight. 3 hours after ingestion of aspirin, the animals were sacrificed. The abdomen was opened and a ligature was placed around the esophagus. The stomach was removed and the contents were drained into graduated centrifuge tube through a small nick along the greater curvature. The volume of the juice was measured.

Induction of gastric ulcer: The animals were fasted for 36 hours in separate cages with raised wide wire mesh to avoid coprophagia (Basso *et al.*, 1983), but with water given *ad libitum*. After 4 hours under light anesthesia by ketamine, animals were killed, stomach was removed and opened on greater curvature and examined for ulceration. Evaluation of the degree of ulceration was expressed in terms of ulcer score which is calculated by dividing the total number of ulcers in each group by a number of rats in that group (Robert *et al.*, 1968). The degree of ulceration was also expressed as ulcer index and calculated by multiplying ulcer score x 100 (Radwan *et al.*, 2003).

Preventive index was calculated according to the method

Preventive index = $\frac{\text{U.I. control} - \text{U.I. treated}}{\text{U.I. control}} \times 100$

$$\% \text{ of ulceration} = \frac{\text{Number of ulcerated rat} - \text{number of non-ulcerated rats} \times 100}{\text{Number of rat in group}}$$

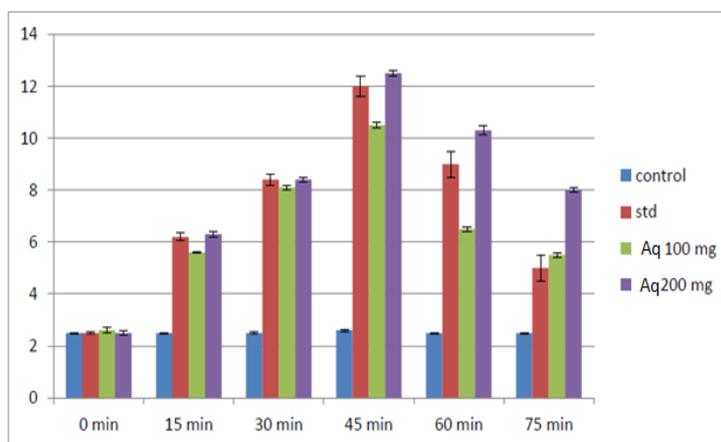
Statistical analysis: The values Mean \pm SEM are calculated for each parameter. For determining the significant intergroup difference each parameter was analyzed separately and one-way analysis of variance^[13] (Gennaro, 1995) was carried out and the individual comparisons of the group mean values were done using Dunnet's test^[14] (Dunnet, 1964).

RESULTS

Table 1: Results Of Aqueous Extract Of *Carum carvi* Seeds By Using Tail Immersion Method

Time in minutes	At 0 min Time in sec	At 15min Time in sec	At 30 min Time in sec	At 45 min Time in sec	At 60 min Time in sec	At 75 min Time in sec
Control	2.5 \pm 0.02	2.5 \pm 0.02	2.55 \pm 0.05	2.6 \pm 0.05	2.5 \pm 0.02	2.5 \pm 0.02
Standard	2.5 \pm 0.05***	6.2 \pm 0.15***	8.4 \pm 0.2***	12 \pm 0.4***	9 \pm 0.5***	5 \pm 0.5***
Aqueous 100mg	2.6 \pm 0.11ns	5.6 \pm 0.04**	8.1 \pm 0.07**	10.5 \pm 0.1**	6.5 \pm 0.09**	5.5 \pm 0.1**
Aqueous 200mg	2.5 \pm 0.1ns	6.3 \pm 0.09ns	8.4 \pm 0.1ns	12.5 \pm 0.1**	10.3 \pm 0.16**	8 \pm 0.08**

Values are mean \pm SD, n=6 rats in each group. Significant values calculated by dunnet's multiple comparison test. Here p values, ***p : <0.001 - Highly significant, **P: < 0.01 – Significant p value > 0.05 – Non significant

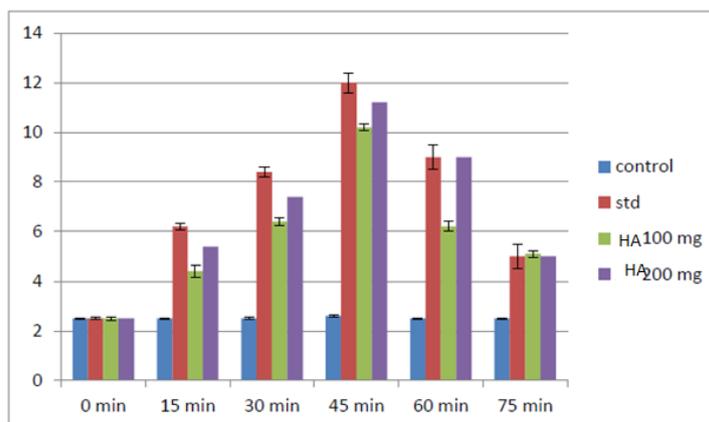


GRAPH 2: Graph Showing Effect of Aqueous Extract On Time In Sec Results By Tail Immersion Method.

Table 2: Results of Hydro alcoholic Extract of *Carum carvi* Seeds by Using Tail Immersion Method.

Time in min	At 0 min Time in sec	At 15 min Time in sec	At 30 min Time in sec	At 45 min Time in sec	At 60 min Time in sec	At 75 min Time in sec
Control	2.5±0.2	2.5±0.2	2.5±0.5	2.6±0.05	2.5±0.02	2.5±0.02
Standard	2.5±0.05***	6.2±0.15***	8.4±0.2***	12±0.4***	9±0.5***	5±0.5***
Hydro alcoholic 100mg	2.5±0.07ns	4.4±0.23**	6.4±0.15**	10.2±0.14**	6.2±0.20**	5.1±0.14ns
Hydro alcoholic 200mg	2.5±0.08ns	5.4±0.09**	7.4±0.15**	11.2±0.17**	9±0.37ns	5±0.1ns

Values are mean±SD, n=6 rats in each group. Significant values calculated by dunnet's multiple comparison test. Here p values, ***p : <0.001 - Highly significant, **P: < 0.01 – Significant p value > 0.05 – Non significant.

**Graph 1: Graph Showing Effect Of hydro alcoholic Extract on Time in Sec Results by Tail Immersion Method.****Table 3: Results Of Anti-Ulcer Activity Using Hydro-Alcoholic Extract Of *Carum carvi* Seeds Against Aspirin-Induced Gastric Ulcers.**

Group	Volume Of Centrifuge(ml)	Volume Of NaOH Consumed(ml)	Total Acidity (mEq/L)	Ulcer Index	% Ulcer Protection
Control	1±0.36	2.5ml±0.72	25±2.56	6±1.05	-
Standard	0.8±0.34	1±0.36	10±0.45***	1.5±0.51***	75±3.57
Hydro alcoholic 100 mg	0.7±0.37	2.1±0.4	21±1.96**	5±0.7**	16.7±0.37
Hydro alcoholic 200 mg	0.4±0.58	1±0.36	10±0.47ns	2±0.58**	66.7±1.09

Values are mean±SD, n=6 rats in each group. Significant values calculated by dunnet's multiple comparison test. Here p values ***p : <0.001 - Highly significant **P: < 0.01 – Significant p value > 0.05 – Nonsignificant

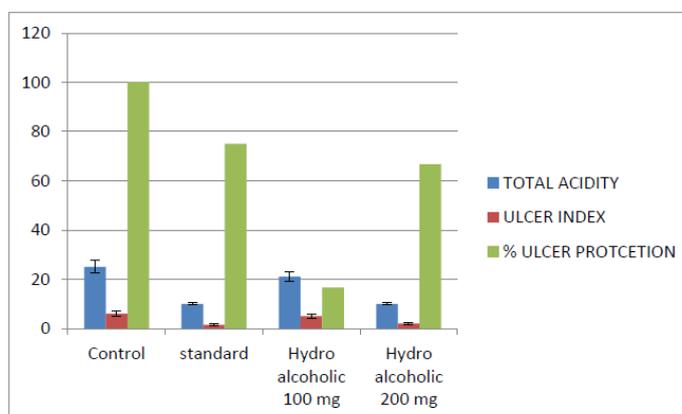
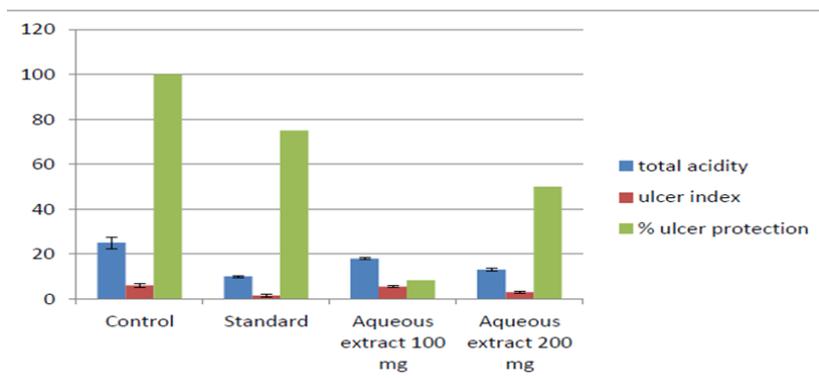
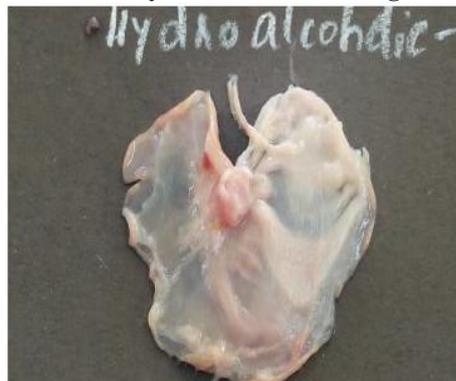
**Graph 3: Graph Showing Effect of Hydroalcoholic Extract on Total Acidity, Ulcer Index, % Ulcer Protection in Rats.**

Table 4: Results Of Anti-Ulcer Activity Using Aqueous Extract of *Carum carvi* Seeds Against Aspirin-Induced Gastric Ulcers.

Group	Volume Of Centrifuge(ml)	Volume Of NaOH Consumed(ml)	Total Acidity (mEq/L)	Ulcer Index	% Ulcer Protection
Control	1±0.36	2.5±0.72	25±2.56	6±1.05	-
Standard	0.8±0.34	1±0.36	10±0.45***	1.5±0.51***	75±3.57
Aqueous extract 100 mg	0.8±0.36	1.8±0.53	18±0.58**	5.5±0.52**	8.3±0.38
Aqueous extract 200 mg	0.4±0.58	1.3±0.36	13±0.55**	3±0.48**	50±0.51

Values are mean±SD, n=6 rats in each group. Significant values calculated by dunnet's multiple comparison test. Here p values ***p : <0.001 - Highly significant **P: < 0.01 – Significant p value > 0.05 – Nonsignificant

**GRAPH 4: Graph Showing Effect Of Hydro Extract On Total Acidity, Ulcer Index, % Ulcer Protection In Rats.****Hydroalcoholic-100mg****Hydro-alcoholic 200 mg****Aqueous – 100mg****Aqueous – 200mg**

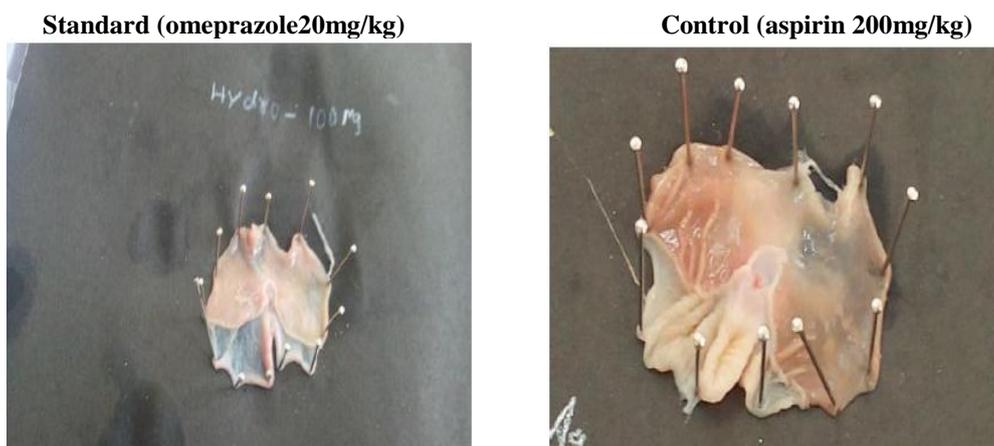


Fig. 8: Images showing ulcer index produced in animals during Screening of Anti-Ulcer Activity of two different solvent extracts of *Carum carvi* against Aspirin-induced gastric ulcers in rats.

Phytochemical Screening Results

Table 2: Table Showing Results Of Phytochemical Screening:

S. no.	Name Of The Experiment	Hydroalcoholic Extract	Hydro Extract
1	Detection Of Alkaloids:	-	-
2	Test For Glycosides:	-	-
3	Test for cardiac glycosides:	-	-
4	Tests for deoxy sugars:	-	-
5	Tests for anthraquinone glycosides:	-	-
6	Tests for cyanogenetic glycosides	-	-
7	Test for saponin glycosides:	-	-
8	Test for coumarin glycosides	-	-
9	Test for carbohydrates:	-	-
10	Test for non-reducing sugars:	-	-

DISCUSSION

Analgesic Studies: The time taken in seconds for the rat to withstand its tail in hot water at 55°C was noted. In control animals, without any drug, almost all rats withstand for approximately 2.5 sec at varying time intervals. The time taken for the standard group i.e., group of rats treated with standard drug Diclofenac sodium the time in seconds for the rat to withdraw its tail from hot water is increased with increase in time. At a maximum of 45 min after ingestion of standard drug the rat could withstand for 12 sec showing the analgesic effect. In comparison with the time in second values at increasing time in test groups treated with hydro (or) aqueous extract 100mg & 200mg and also in the group treated with a hydroalcoholic extract of 100mg & 200mg, the values are also increasing.

Determination of Ulcer Index: The mean ulcer index value in control group is 6 and for standard drug Omeprazole. The mean ulcer index for test groups treated with hydroalcoholic extract 100mg & 200mg is 5±0.7 & 2±0.58 and in group treated with hydro/aqueous extract at dose of 100mg & 200mg is 5.5±0.52 & 3±0.48.

Determination of Percentage Ulcer Protection: In the control group, almost all animals are positive for an ulcer. For the group treated with standard drug

Omeprazole 75% of the population in the group is protected from an ulcer. In the case of groups treated with a hydroalcoholic extract of 100mg & 200mg dose almost 16.7% & 66.7% protection is observed in a population and in the case of the group treated with aqueous extract at a dose of 100mg and 200 mg 8.3% and 50% protection is observed.

Phytochemical screening: proteins, tannins and phenolic compounds, flavonoids are present. Alkaloids, glycosides, carbohydrates, non-reducing sugars, steroids, amino acids, saponins, volatile oils are absent in the screening of phytochemicals.

CONCLUSION

Review literature supported the use of seeds of *Carum carvi* for Analgesic and Anti-Ulcer Activity. Extraction using aqueous and hydroalcoholic solvents was done. Screening of Analgesic activity is done by using tail immersion method at a dose of 100 mg and 200 mg by using two extracts. In comparison with the standard drug, the results of hydroalcoholic extract at 100 mg dose showed good analgesic activity compared with a standard drug. Anti-Ulcer activity is screened by using Aspirin-induced ulcer at doses of 100 mg and 200 mg. At a dose of 200mg of hydroalcoholic extract, the percentage of ulcer protection in test group is almost similar with ulcer protection with a standard drug. It

decreases the total acidity and ulcer index. Finally, we conclude that hydroalcoholic extract at a dose of 100 and 200mg shows good Analgesic and Anti-Ulcer Activity. The drug was selected for a future investigation involving column chromatographic separation of extracts and to isolate and identify the actual chemical constituent responsible for the activity.

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