



**REM SLEEP DEPRIVATION-INDUCED OXIDATIVE STRESS AND ITS  
ATTENUATION BY *WITHANIA SOMNIFERA* (L.) IN DISCRETE REGIONS OF RAT  
BRAIN.**

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**ABSTRACT**

Currently, plants are used as therapeutic agents in the wide range of clinical applications. The aim of this study is to assess the Antioxidant activity of *Withania somnifera* in 72 hours REM sleep deprivation. The plant (root) sample collected and Ethanolic extraction was done by using Soxhlet apparatus. Five groups of Wistar strain male albino rats were used in this study. Each group comprises of 6 rats and multiple platform models used for REM sleep deprivation. The values were statistically analyzed using one-way analysis of variance (ANOVA) followed by Tukey's posthoc test for multiple comparison methods. The Significance level was kept at P<0.05. The activity of Superoxide dismutase, Lipid peroxidation, Catalase, Glutathione peroxidase, levels of reduced glutathione, Vitamin C, Vitamin E in discrete regions of the rat brain and plasma Corticosterone level were measured. *Withania somnifera* possesses remarkable antioxidant activity by normalizing all the altered enzymes and plasma corticosterone level as a result of 72 hours REM sleep deprivation exposure.

**KEYWORDS:** Antioxidants, *Withania somnifera*, REM sleep deprivation, oxidative stress and corticosterone.

**INTRODUCTION**

Sleep is essential for normal physiological functions in human and animals; it provides restorations of emotional and physical activity in a manner that is not well established.<sup>[1]</sup> The average sleep per 24 hours has declined by 1.5 hours over the past century.<sup>[2]</sup> Sleep occurs in two stages: Non rapid eye movement (NREM) and rapid eye movement (REM).<sup>[3]</sup> Evidence from many studies suggests that an important function of sleep is a consolidation of new information into long-term memory. Multiple studies in both humans and rodents reported memory impairments as a result of sleep deprivation (SD).<sup>[4]</sup> Human studies show that the total SD for a single night disturbs memory function.<sup>[5]</sup> Additionally, numerous animal studies also shown that 24 hours to 96 hours of SD lead to impairment of memory and behavioral changes. Moreover, SD was shown to impair long-term potentiation of the hippocampus, which is one of the main centers for learning and memory function.<sup>[6]</sup> SD also elevates hippocampal oxidative stress, which reflects on neuronal excitability, cognitive functions and molecular signaling.<sup>[7]</sup> SD reduces the expression of transcription

and translation synaptic proteins in hippocampus and many regions of the brain.<sup>[8]</sup> During pregnancy SD increases the risk of preeclampsia, gestational diabetes, intrauterine growth restriction and the need for cesarean delivery.<sup>[9]</sup> Cytokines like Tumor necrosis factor-alpha and Interleukin 1- beta gene expression increases in the hypothalamus during REM sleep deprivation.<sup>[10]</sup> SD leads to stroke, obesity, diabetes, osteoporosis, cardiovascular disease, cancer and permanent cognitive deficits.<sup>[11]</sup>

*Withania somnifera* (WS) known commonly as ashwagandha and belongs to Solanaceae family, the green shrub plant has been used in India as traditional medicine for the treatment of mental health, ageing process<sup>[12]</sup> and promote learning and memory in animal models.<sup>[13]</sup> It also has antioxidant activity, according to the Indian indigenous system of medicine. Hence this present study was focused to evaluate physiological antioxidant defense elements, like lipid peroxidation (LPO), superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), reduced glutathione

(GSH), Vitamin C and Vitamin E in discrete regions of rat brain.

## MATERIALS AND METHODS

### Animals

Wistar strain male albino rats (200- 250g) were maintained under standard laboratory conditions with water and food, the animals were handled according to the principles of laboratory care framed by the committee for the purpose of control and supervision of experiments on animals (CPCSEA), Government of India. Animal experiments were carried out after getting clearance from the institutional animal ethical committee (IAEC NO: 55/12/2015).

### Experimental protocol

Animals were divided into five equal groups; each group consists of six animals. The Ethanolic extract of *Withania somnifera* was used for this study.

Group - I Control animals were used for studying the baseline values.

Group - II Animals were administered 2% of Tween 80 as a vehicle.

Group - III Animals were exposed to 72 hours REM sleep deprivation.

Group - IV Animals were treated with only an Ethanolic extract of *Withania somnifera* (500 mg/kg) <sup>[14]</sup> for 18 days and

Group - V Animals pretreated with Ethanolic extract of *Withania somnifera* for 15 days +72 hours REM sleep deprivation with the same treatment.

### Plant collection and extract preparation

The dried root powder of *Withania somnifera* was purchased from Government Irua Tribe Women's Welfare Society (ITWWS), Tandarai, Chengalpattu, Tamilnadu, India- 603001. Bill No: 130/05/2016. About 200 g of root powder was extracted with 1000 ml of 95% Ethanol under reflux by heating over a water bath at 60°C. The extract was then vacuum dried. The yield of Ethanolic extract of *Withania somnifera* was 31.23 % (w/w). The suspension of the extracted drug was prepared by dissolving in 2% Tween 80 before administration to animals.<sup>[15]</sup>

### REM sleep deprivation

REM sleep deprivation (RSD) for 72 hrs was induced by using the multiple platform models.<sup>[16]</sup> It was started and ended at the beginning of the light phase and the room was maintained at the controlled temperature (23±1° C) and light-dark cycle (lights on between 07.00 and 19.00 hrs). In this experiment 6 rats from same group placed in a water tank (120 cm x 70cm x 50 cm) containing 10 round platforms (each round platform made up of 7 cm diameter and 10 cm height, it was raised 2 cm above the

water level) arranged in two lines and 20 cm away from each other (edge to edge), in which the rats can move around freely from one platform to another one. Loss of muscle tone at the beginning of each REM (paradoxical) sleep episode causes rats to fall in the water, thus being awakened. During the SD period, the animals had free access to water bottles and chow pellets attaching from a grill located on the top of the chamber. As the animals can move freely within the multiplatform chamber, it has been reported that it has less immobilization stress compared to the single version of platform technique.<sup>[17]</sup>

### Biochemical analysis

The activity of LPO was indirectly estimated by determining the accumulation of thiobarbituric acid reactive substances (TBARS) in the tissue homogenate by the method of Ohkawa *et al.*<sup>[18]</sup> According to Markland and Markland<sup>[19]</sup> the SOD activity was measured as the degree of inhibition of auto-oxidation of pyrogallol at alkaline pH. The activity of catalase was measured as the amount of hydrogen peroxide consumed per minute per milligram of protein by the method of Sinha.<sup>[20]</sup> Glutathione peroxidase level was estimated by measuring the amount of reduced glutathione consumed in the reaction mixture according to the method of Rotruck *et al.*<sup>[21]</sup> The reduced glutathione level was measured based on the development of relatively stable yellow color, when 200mM 5, 5' - dithiobis- (2-nitrobenzoic acid) (DTNB) solution was added, according to the method of Moron *et al.*<sup>[22]</sup> Vitamin C was estimated by the method of Omaye *et al.*<sup>[23]</sup> and Vitamin E was measured by the method of Desai.<sup>[24]</sup> The total protein was estimated by the method of Lowry *et al.*<sup>[25]</sup> using Bovine serum albumin (BSA) as standard. The corticosterone level was measured by using Elisa kit. (Cusabio, biotech Co Limited, China. C.No.CSB-E07014r).

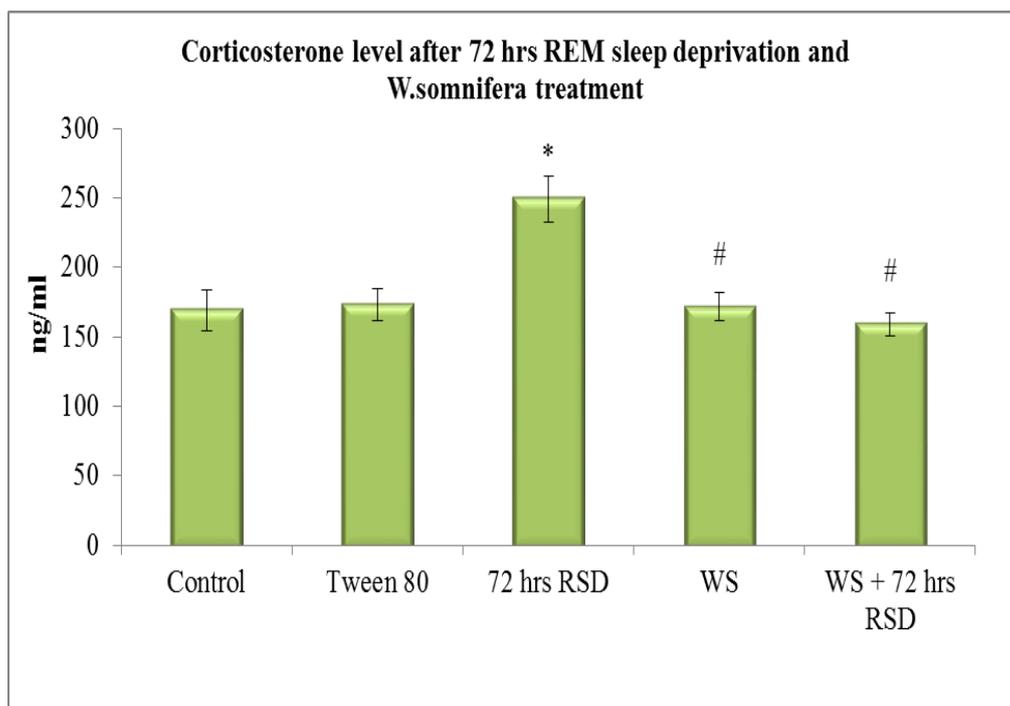
### Statistical analysis

All data were expressed as Mean ± Standard deviation (STDEV). The statistical significance was evaluated by one - way analysis of variance (ANOVA) using SPSS statistical package version 20.0 (SPSS, Cary, NC, USA). When there is a significant difference, Tukey's multiple comparison tests were performed by fixing the significance at level  $P < 0.05$ .

## RESULTS AND DISCUSSION

### Plasma Corticosterone

The plasma corticosterone (d.f.=4, F=43.131) levels "Fig.1" were increased in Group-III compared to Group-I control. However, corticosterone levels in Group-V animals were significantly decreased when compared to Group-III after 72 hours REM sleep deprivation exposure.

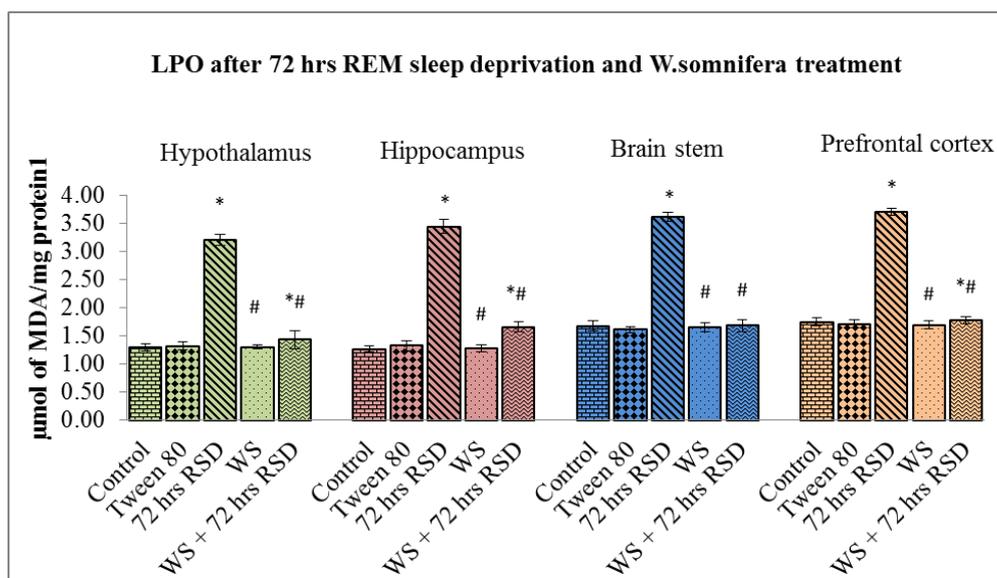


**Fig. 1:** Data are expressed as Mean  $\pm$  STDEV. for six rats in each group. The values are expressed ng/ml. \* Compared with Control; # Compared with 72 hrs REM sleep deprivation. The symbols represent statistical significance: \*, #  $P < 0.05$ .

#### Lipid peroxidation

The levels of LPO "Fig. 2" were significantly increased in Group-III after 72 hours REM sleep deprivation in discrete regions of the brain [Hypothalamus (d.f.=4,  $F=461.326$ ), Hippocampus (d.f.=4,  $F=655.724$ ), Brainstem (d.f.=4,  $F=667.603$ ) and Prefrontal

cortex(d.f.=4,  $F=1047.254$ )] when compared to Group-I control (animals not exposed to REM sleep deprivation). The LPO levels were lower in the drug-treated group (Group-V) compared to the Group-III animals. The values of decreased LPO level in Group-V were not statistically different from Group-I animals.



**Fig. 2:** Data are expressed as Mean  $\pm$  STDEV for six rats in each group. The values are expressed  $\mu\text{mol}$  of MDA formed  $\text{mg protein}^{-1}$ . \* Compared with Control; # Compared with 72 hrs REM sleep deprivation. The symbols represent statistical significance: \*, #  $P < 0.05$ .

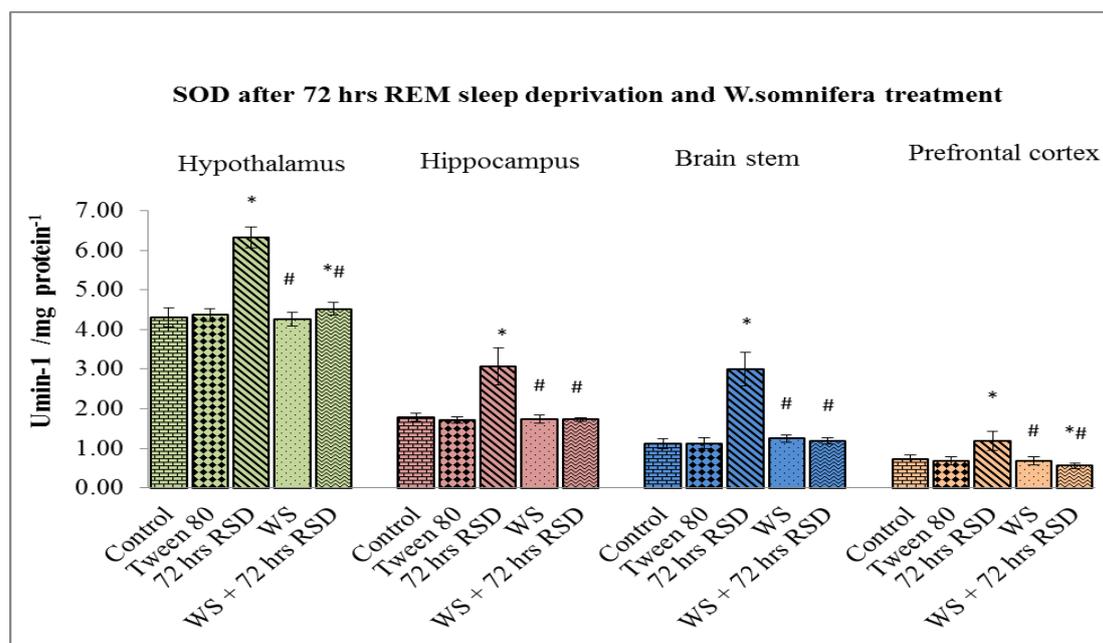
#### Superoxide dismutase

The SOD is not evenly distributed in the regions studied as indicated by the control values. The activities of SOD

"Fig. 3" were significantly increased in Group-III after 72 hours REM sleep deprivation in discrete regions of the brain [Hypothalamus (d.f.=4,  $F=108.284$ ),

Hippocampus (d.f.=4, F=41.392), Brainstem (d.f.=4, F=86.756) and Prefrontal cortex(d.f.=4, F=17.374)] when compared to Group-I. The SOD activities were lower in the drug-treated group (Group-V) compared to

the Group-III animals. The activities of SOD decreased in Group-V were not statistically different from Group-I animals.

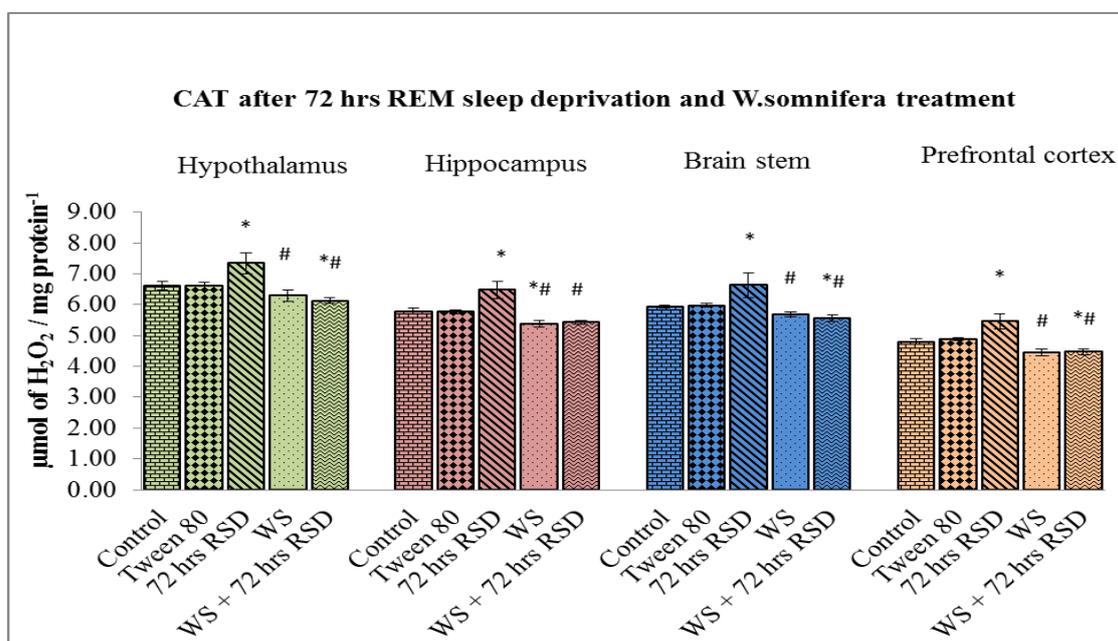


**Fig. 3:** Data are expressed as Mean  $\pm$  STDEV for six rats in each group. The values are expressed Umin<sup>-1</sup> mg protein<sup>-1</sup>. \* Compared with Control; # Compared with 72 hrs REM sleep deprivation. The symbols represent statistical significance: \*, # P < 0.05.

#### Catalase

The results are summarized in "Fig. 4". The catalase levels were increased in Group-III animals after 72 hours REM sleep deprivation in discrete regions of the brain [Hypothalamus (d.f.=4, F=35.736), Hippocampus

(d.f.=4, F=52.934), Brainstem (d.f.=4, F=27.985) and Prefrontal cortex (d.f.=4, F=52.783)] when compared to Group-I. The catalase levels were lower in the drug-treated group (Group-V) compared to the Group III animals.

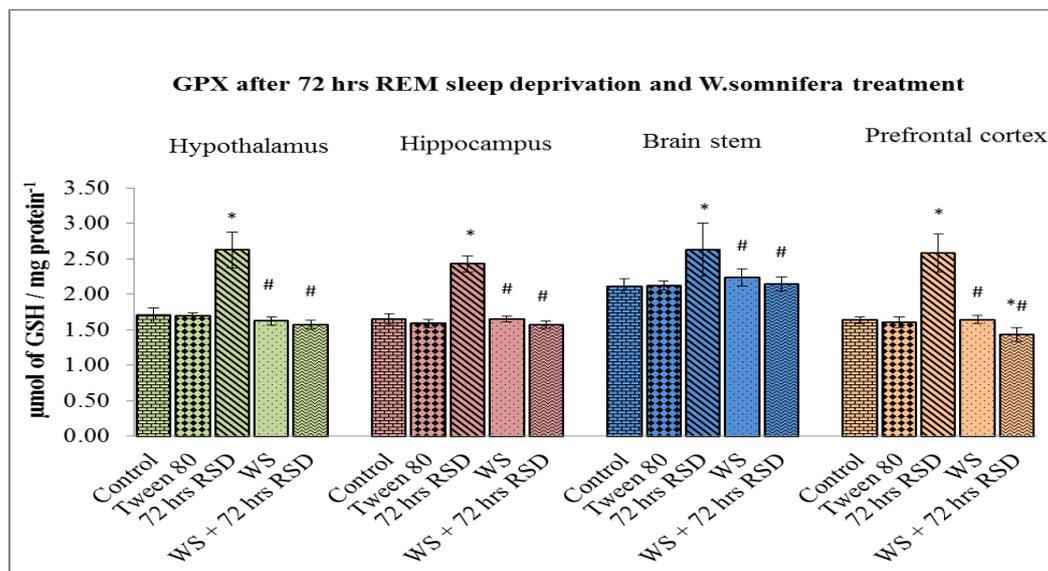


**Fig. 4:** Data are expressed as mean  $\pm$  STDEV for six rats in each group. The values are expressed  $\mu\text{mol of H}_2\text{O}_2$  consumed min<sup>-1</sup> mg protein<sup>-1</sup>. \* Compared with Control; # Compared with 72 hrs REM sleep deprivation. The symbols represent statistical significance: \*, # P < 0.05.

**Glutathione peroxidase**

The results are summarized in "Fig. 5". The GPx also showed regional variations among the brain region studied. The GPx levels increased in Group-III after 72 hours REM sleep deprivation in discrete regions of the brain [Hypothalamus (d.f.=4, F=71.217), Hippocampus

(d.f.=4, F=150.491), Brainstem (d.f.=4, F=7.066), and Prefrontal cortex(d.f.=4, F=66.037)] when compared to Group-I. The GPx levels were lower in the drug-treated group (Group-V) compared to the Group-III animals. The values of decreased GPx in Group- V were not statistically different from Group – I animals.

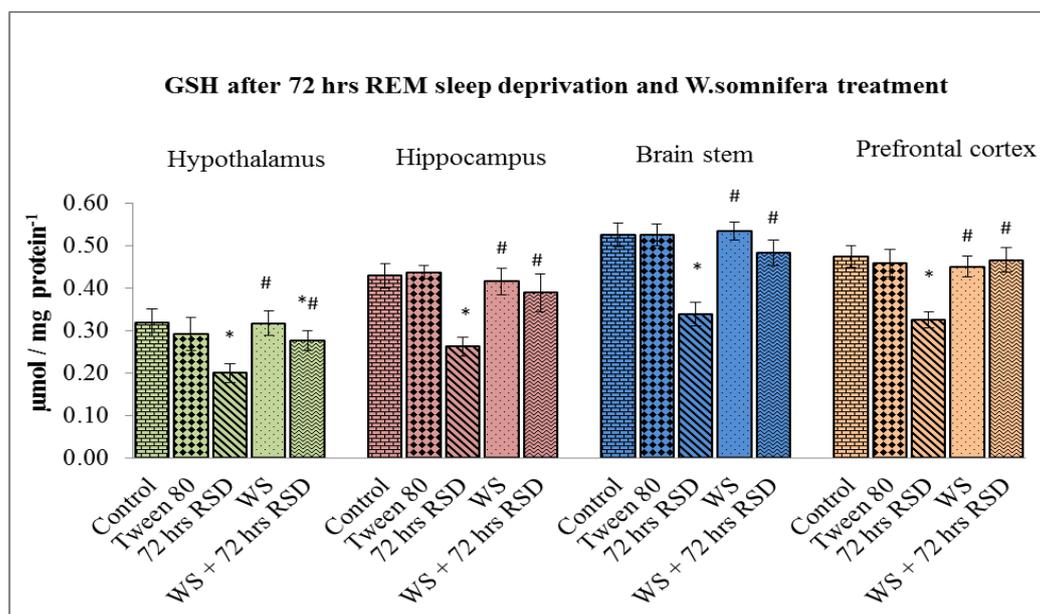


**Fig. 5:** Data are expressed as Mean  $\pm$  STDEV for six rats in each group. The values are Expressed  $\mu\text{mol}$  of GSH oxidized  $\text{min}^{-1} \text{mg protein}^{-1}$ . \* Compared with Control; # Compared with 72 hrs REM sleep deprivation. The symbols represent statistical significance: \*, #  $P < 0.05$ .

**Reduced glutathione**

The results are summarized in "Fig. 6". The GSH levels were significantly decreased in Group - III animals after 72 hours of REM sleep deprivation in discrete regions of the brain [Hypothalamus (d.f.=4, F=15.617), Hippocampus (d.f.=4, F=32.587), Brainstem (d.f.=4,

F=51.122), and Prefrontal cortex (d.f.=4, F=27.979)] when compared to Group-I animals. The GSH levels were higher in the drug-treated group (Group-V) compared to the Group-III animals. The values of increased GSH in Group-V were not statistically different from Group-I animals.



**Fig. 6:** Data are expressed as Mean  $\pm$  STDEV for six rats in each group. The values are expressed  $\mu\text{mol}$   $\text{mg protein}^{-1}$ . \* Compared with Control; # Compared with 72 hrs REM sleep deprivation. The symbols represent statistical significance: \*, #  $P < 0.05$ .

**Vitamin C and E**

Vitamin C (Table.1) and Vitamin E (Table.2) levels were significantly decreased in Group - III animals after 72 hours REM sleep deprivation when compared to Group-I Control animals. On the other hand, Vitamin C and

Vitamin E levels were higher in the drug-treated group (Group- V), compared to Group-III animals. The values of increased Vitamin C and Vitamin E in Group - V were not statistically different from Group - I animals.

**Vitamin C Level after 72 hrs REM sleep deprivation and *W.somnifera* treatment**

Brain regions	Group 1	Group 2	Group 3	Group 4	Group 5
Hypothalamus (d.f.=4, F=315.124)	339.50±8.30	338.88±5.12	232.56±6.48 *	345.70±5.88 #	333.61±7.05#
Hippocampus (d.f.=4,F=152.482)	394.50±4.02	389.30±5.82	287.25±11.06 *	381.50±10.17 #	370.98±10.57*#
Brain stem (d.f.=4,F=226.853)	292.31±4.98	291.90±5.39	220.48±5.68 *	286.25±4.99#	275.51±3.66*#
Pre frontal cortex (d.f.=4,F=284.817)	485.16±6.35	479.58±12.47	327.21±5.73 *	475.50±13.47#	460.95±7.47*#

**Table 1. Data are expressed as Mean ± STDEV for six rats in each group. The values are expressed µg /g tissue<sup>-1</sup>. \* Compared with Control; # Compared with**

**72 hrs REM sleep deprivation. The symbols represent statistical significance: \*, # P <0.05.**

**Vitamin E Level after 72 hrs REM sleep deprivation and *W.somnifera* treatment.**

Brain regions	Group 1	Group 2	Group 3	Group 4	Group 5
Hypothalamus (d.f.=4,F=21.758)	11.85±1.16	11.72±0.82	7.79±0.70*	11.14±0.51#	12.25±0.95#
Hippocampus (d.f.=4,F=47.613)	16.52±0.73	17.36±0.68	12.25±0.72*	16.25±1.33#	16.12±0.87#
Brain stem (d.f.=4,F=79.482)	15.82±0.93	15.31±0.94	8.66±0.42*	13.33±0.60*#	12.82±0.53*#
Pre frontal cortex (d.f.=4,F=69.133)	17.90±0.88	18.73±0.68	12.44±0.70*	16.56±0.40*#	14.63±1.06*#

**Table 2. Data are expressed as Mean ± STDEV for six rats in each group. The values are expressed µg /g tissue<sup>-1</sup>. \* Compared with Control; # Compared with 72 hrs REM sleep deprivation. The symbols represent statistical significance: \*, # P <0.05.**

In Group-II and IV animals, mostly all the parameters in all the regions of the brain were not significantly different from Group-I animals.

Many studies have shown that SD-induced oxidative damage in several types of tissues.<sup>[26]</sup> One of the studies that Sleep deprivation alters gene expression and antioxidant enzyme activity in mice.<sup>[27]</sup> Oxidative stress had linked with cognitive impairments.<sup>[28]</sup>

Antioxidants and free radicals scavenging system present in the cell protect against the damaging effects of free radicals.<sup>[29]</sup> In normal cells, there is a balance exists between oxidative products and antioxidants, like SOD, Catalase, GPx, GSH, Vitamin C and Vitamin E. Brain is more vulnerable to oxidative damage due to its high oxygen consumption.<sup>[30]</sup> For the transmission of impulses between the neuronal cells, it requires energy in the form of ATP, during the production of ATP using oxygen, a small percentage (less than 3%) of oxygen in mitochondria is mindfully converted to superoxide radical (SOR) This explains the reason for the increased

production of free radicals in the brain. Also stress response results in the creation of some other reactive oxygen species such as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hydroxyl radicals (. OH) and superoxide anion radicals (O<sub>2</sub><sup>-</sup>) that cause lipid peroxidation, especially in cell membrane leads to tissue damage.<sup>[31]</sup> The platform technique used for SD raises the possibility that long-term impairment of synaptic plasticity.<sup>[32]</sup>

In the present study, 72 hours REM sleep deprivation are producing marked and increased LPO, SOD, CAT, GPx activities and decreased in the levels of GSH, Vitamin C and Vitamin E in discrete regions of the brain that were tested (Hypothalamus, Hippocampus, Brainstem and Prefrontal cortex). Further, it also significantly increases the Corticosterone level in plasma.

Acute exposure to stress leads to an excessive generation of oxygen free radicals this might be the reason for the production of the excess amount of LPO.<sup>[33]</sup> SOD is a major intracellular enzyme, which protects the cell against oxygen free radicals by speeding up its dismutation of the superoxide anion (O<sub>2</sub><sup>-</sup>). In recent study states that the redox-sensitive transcription factor NF-E2-related factor 2 (Nrf2) plays an important role in activating antioxidant enzymes, the increased SOD activity could be due to the activation of Nrf2 induced by SD.<sup>[34]</sup> Catalase was responsible for the detoxification of

significant amounts of H<sub>2</sub>O<sub>2</sub>. So, catalase level was increased in 72 hour REM sleep deprivation.

GPx is a major antioxidant enzyme in many tissues, especially in the brain, which metabolizes peroxides, such as H<sub>2</sub>O<sub>2</sub> and protects cell membranes from lipid peroxidation.<sup>[35]</sup> GPx activity in the brain is more important than CAT for the destruction of H<sub>2</sub>O<sub>2</sub> because this enzyme was more in mitochondria and cytosol. The GSH levels were significantly reduced in SD animals; this might be due to its multiple activities in scavenging the free radicals produced by SD, which leads to its greater consumption.<sup>[36]</sup> In the present study also the GSH level was significantly reduced in the 72 hour SD.

Vitamin C acts as a powerful scavenger of superoxide – induced lipid peroxidation. Endogenous Vitamin C and Vitamin E levels have been reported to decline under stress conditions.<sup>[37]</sup> Plants have both enzymatic and non-enzymatic systems to scavenge active oxygen species. Vitamin C and E are non enzymatic compounds can protect the human body from free radicals.<sup>[38]</sup>

Vitamin E is a powerful antioxidant that inhibits free radical reaction and prevents oxidative stress.<sup>[39]</sup> Further, Vitamin E administration to an animal model of diabetes normalizes the GSH level and activities of GPx, Catalase and SOD.<sup>[40]</sup>

In the present study, *Withania somnifera* treated sleep-deprived animals showed a marked decrease in SOD, GPx and Catalase activity when compared with sleep-deprived animals. The further LPO level was also significantly decreased in the *Withania somnifera* treated sleep-deprived animals. GSH, Vitamin C and Vitamin E level were significantly increased in the *Withania somnifera* treated sleep-deprived animals when compared with the sleep-deprived animals.

The roots of *Withania somnifera* contain the Withanolides (withanolide A and 12-deoxy withastramonolide) and withaferin A.<sup>[41]</sup> Also, it has male infertility activity,<sup>[42]</sup> the neuroprotective effect,<sup>[43]</sup> hypoglycemic activity,<sup>[44]</sup> and an antibacterial and antimutagenic activity.<sup>[45]</sup>

## CONCLUSION

The observed data confirmed the presence of oxidative damage in the discrete regions of the brain when exposed to 72 hours REM sleep deprivation. The significant variations in SOD, LPO, CAT, GPx, GSH, Vitamin C, Vitamin E and corticosterone were normalized by an Ethanolic extract of *Withania somnifera* in the rat. Further, there was no adverse effect found in the rat model of treatment with *Withania somnifera* alone. In future, it is necessary to conduct more extensive studies with *Withania somnifera* to explicate their molecular mechanism in antioxidant activity.

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## Authors contributions

All the authors have contributed equally.

## Disclosure of Interests

The authors declare that they have no conflicts of interest concerning this research article.

## REFERENCES

1. Siegel JM. Sleep viewed as a state of adaptive inactivity. *Nat Rev Neurosci*, 2009; 10: 747.
2. Ramanathan L, Gulyan S, Nienhuis R, *et al.* Sleep deprivation decreases superoxide dismutase activity in rat hippocampus and brain stem. *Neuroreport*, 2002; 13: 1387–1390.
3. Guyton and Hall, *Textbook of Medical Physiology*. Elsevier Saunders publications, International 11<sup>th</sup> edition, 2006; 739-741.
4. Walker MP, Stickgold R. Sleep-dependent learning and memory consolidation. *Neuron*, 2004; 44: 121–133.
5. VanderWerf YD, Altena E, Schoonheim MM, Sanz-Arigita EJ, *et al.* Sleep benefits subsequent hippocampal functioning. *Nat Neurosci*, 2009; 12: 122–123.
6. Aleisa AM, Helal G, Alhaider IA, Alzoubi KH, *et al.* Acute nicotine treatment prevents REM sleep deprivation-induced learning and memory impairment in the rat. *Hippocampus*, 2011; 21: 899–909.
7. Alhaider IA, Aleisa AM, Tran TT, Alzoubi KH, Alkadhi KA. Chronic caffeine treatment Prevents sleep deprivation-induced impairment of cognitive function and synaptic plasticity. *J Sleep*, 2010; 33: 437-44.
8. Porter NM, Bohannon JH, Curran-Rauhut M, Buechel HM, *et al.* Hippocampal CA1 transcriptional profile of sleep deprivation: relation to aging and stress. *PLoS One*, 2012; 7: e40128.
9. Hutchison BL, Stone PR, McCowan LM, *et al.* A postal survey of maternal sleep in later pregnancy. *BMC Pregnancy, Childbirth*, 2012; 12: 144.
10. Won Sub Kanga, Hae Jeong Parkb, Joo-Ho Chungb, Jong Woo Kima. REM sleep deprivation increases the expression of interleukin genes in mice hypothalamus. *Neuroscience Letters*, 2013; 556: 73–78.
11. Colten HR, Altevogt BM. Institute of Medicine (US) Committee on Sleep Medicine and Research. *Sleep disorders and sleep deprivation: an unmet public health problem. Extent and health consequences of chronic sleep loss and sleep disorders*, vol. 3. Washington, DC: National Academies Press (US); 2006. Available at: <http://www.ncbi.nlm.nih.gov/books/NBK19961/>.
12. Bhattacharya SK, Bhattacharya D, Sairam K, Ghosal S. Effect of *Withania somnifera* glycowithanolides

- on a rat model of tardive dyskinesia. *Phytomedicine*, 2002; 9: 167–170.
13. Dhuley JN, Nootropic like effect of Ashwagandha (*Withania somnifera*) in mice. *Phytother. Res*, 2001; 15: 524–528.
  14. Shruti B, Nirav J, Lal I. Safety assessment of *Withania somnifera* extract standardized for withaferin A: Acute and sun- acute toxicity study. *Journal of ayurveda and integrative medicine*, 2016; 7: 30–37.
  15. Singh, Sukhwinder, Anu. Effect of ethanolic extract of *Withania somnifera* roots on antioxidant defence in mercury induced toxicity in HepG2 cell line. *Online Journal of Pharmacology and Pharmacokinetics*, 2009; 5: 65–72.
  16. Hajali V, Sheibani V, *et al.* Female rats are more susceptible to the deleterious effects of paradoxical sleep deprivation on cognitive performance. *Behavioral Brain Research*, 2012; 228(2): 311–318.
  17. Machado R, Hipolide DC, *et al.* Sleep deprivation induced by the modified multiple platform technique: Quantification of sleep loss and recovery. *Brain Research*, 2004; 1004(1–2): 45–51.
  18. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxidation in animal tissue by the thiobarbituric acid reaction. *Anal Biochem*, 1979; 95: 351–8.
  19. Marklund S, Marklund G. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *EUR J Biochem*, 1974; 47: 469–74.
  20. Sinha AK. Calorimetric assay of catalase. *Anal Biochem*, 1972; 47: 389–94.
  21. Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium biochemical role as a component of glutathione peroxidase. *Science*, 1973; 179: 588–90.
  22. Moron MS, Depierre JW, Mannervik B. Levels of glutathione, glutathione reductase and glutathione-S-transferase activities in rat lung and liver. *Biochim Biophys Acta*, 1979; 582: 67–78.
  23. Omaye ST, Turnbull JD, Sauberlich HE. Selected methods for the determination of ascorbic acid in animal cells, tissues, and fluids. *Methods Enzymol*, 1979; 62: 3–11.
  24. Desai ID. Vitamin E analysis method for animal tissues. *Methods Enzymol*, 1984; 105: 138–43.
  25. Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin-phenol reagent. *J Biol Chem*, 1951; 193: 265–75.
  26. Everson CA, Laatsch CD, Hogg N. Antioxidant defense responses to sleep loss and sleep recovery. *Am J Physiol Regul Integr Comp Physiol*, 2005; 288: R374–83.
  27. Lungato L, Marques MS, Pereira VG, Gazarini ML, Tufik S, Almeida VD. Sleep deprivation alters gene expression and antioxidant enzyme activity in mice splenocytes. *Scandinavian Journal of Immunology*, 2013; 77: 195–199.
  28. Butterfield DA, Drake J, Pocernich C, Castegna A. Evidence of oxidative damage in Alzheimer's disease brain: The central role for amyloid beta-peptide. *Trends Mol MED*, 2001; 7: 548–54.
  29. Naritsara T, Pannapa P, Pattra S. Evaluation of antioxidant and antibacterial activities of fresh and freeze-dried selected fruit juices. *Asian J Pharm Clin Res.*, 2017; 10(9): 156–60.
  30. Floyd, R.A., Carney, J.M. Free radical damage to protein and DNA: Mechanism involved and relevant observation on brain undergoing oxidative stress. *Ann Neurol*, 1992; 32(Suppl.): S22–S27.
  31. Beckman K, Ames B. The free radical theory of aging matures. *Physiol Rev*, 1998; 78: 548–581.
  32. McEwen BS. The neurobiology of stress: from serendipity to clinical relevance. *Brain Res*, 2000; 886: 172–189.
  33. Aravind KN, Mathangi DC, Namasivayam A. Noise-induced changes in free radical scavenging enzymes in the blood and brain of albino rats. *Med Sci Res*, 1998; 26: 811–812.
  34. Na HK, Kim EH, Jung JH, Lee HH, Hyun JW, Surh YJ. Epigallocatechin gallate induces Nrf2-mediated antioxidant enzyme expression via activation of PI3K and ERK in human mammary epithelial cells. *Arch Biochem Biophys*, 2008; 476: 171–7.
  35. Barlow Walden LR, Reiter RJ, Abe M. Melatonin stimulates brain glutathione peroxidase activity. *Neurochem Int*, 1995; 26: 497–502.
  36. Sokolovsk VV, Goncharova LL, Kiseliva NN, Maka Radionova LP. The antioxidant system noise induced stress. *J Med Khim*, 1987; 33(6): 111–113.
  37. Acharya S, Acharya UR. In vivo lipid peroxidation responses of tissues in lead-treated Swiss mice. *Ind Health*, 1997; 35: 542–4.
  38. Pradeesh S, Swapna TS. Antioxidant activity in leaves of *Sesbania grandiflora*(L.) Pers. *Asian J Pharm Clin Res*, 2018; 11(1): 116–119.
  39. Packer L. Vitamin E is nature's master antioxidant. *Sci Am*, 1994; 1: 54–63.
  40. Tiwari V, Kuhad A, Bishnoi M, Chopra K. Chronic treatment with tocotrienol, an isoform of vitamin E, prevents intracerebroventricular streptozotocin-induced cognitive impairment and oxidative-nitrosative stress in rats. *Pharmacol Biochem Behav*, 2009; 93: 183–9.
  41. Tushar Dhanani, Sonal Shah, Gajbhiye, Satyanshu Kumar. Effect of extraction methods on yield, phytochemical constituents and antioxidant activity of *Withania somnifera*. *Arabian Journal of Chemistry*, 2017; 10: S1193–S1199.
  42. Pallav Sengupta, Ashok Agarwal, Maria Pogrebetskaya, Shubhadeep Roychoudhury, Damayanthi Durairajanayagam, Ralf Henkel. Role of *Withania somnifera* in the management of male infertility. *J Reproductive BioMedicine Online March*, 2018; 36(3): 311–326.
  43. Anjali pandey, sarang bani, prabhu dutt, naresh kumar, Krishna avathar, Ghulam nabiqazi. Multifunctional neuroprotective effect of Withanone, a compound from *Withania somnifera*

- roots in alleviating cognitive dysfunction. *J Cytokine*, 2018; 102: 211-221.
44. Gorelic jonathan, Rosenberg rivika, Smotrich Avinoam, Nirit et al. Hypoglycemic activity of withanolides and elicited *Withania somnifera*. *J phytochemistry*, 2015; 116: 283-289.
45. Hemashekhar, Govindappa, Nagaraju, Ganganagappa, Chandrasekar, et al. Green alloy of silver nanoparticles from endophytic extracts of *Withania somnifera* and studies of Antibacterial and antimutagenic activity. *Asian J Pharm Clin Res*, 2017; 10(11): 300-303.