



IN VITRO ANTIOXIDANT ACTIVITY OF *MADHUCA INDICA L.*

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Article Received on 09/04/2018

Article Revised on 29/04/2018

Article Accepted on 19/05/2018

ABSTRACT

The present investigation was aimed to evaluate the phytochemicals and *in vitro* antioxidant activity of different parts of *Madhuca indica*. Qualitative phytochemicals analysis confirmed the presence of carbohydrates, tannins, flavonoids, quinones, glycosides, terpenoids, phlobatannins, alkaloids, phlobatannins, anthraquinones, coumarins, steroids and phytosteroids and absence of phenol, saponins and triterpenoids. *In vitro* antioxidant property was also carried out at five different concentrations (50, 100, 150, 200 and 250 µg/ml) of wood, bark and leaf by total antioxidant capacity, DPPH scavenging activity, hydrogen peroxide scavenging activity and reducing power assay. All the results were compared with standard ascorbic acid. Results showed that dose dependent antioxidant activity in all models. Among the three different parts, bark extract exhibited highest antioxidant activity followed by leaf and wood extract. Our result suggests that bark extract of *Madhuca indica* is a potential source of antioxidant which can be used to treat many free radical mediated diseases.

KEYWORDS: Antioxidant, bark, leaf, *Madhuca indica*, phytochemicals, wood.

INTRODUCTION

Free radicals are defined as molecules having an unpaired electron in the outer orbit. They are generally unstable and very reactive.^[1] Free radicals may play an important role in the origin of life and biological evolution, implicating their beneficial effects on the organisms.^[2] Majority of diseases like atherosclerosis, hyper tension, ischemic disease, alzheimer's disease, parkinsonism, cancer, diabetes mellitus and inflammatory conditions are caused by free radicals which are being considered to be primarily due to the imbalance between pro oxidant and antioxidant homeostasis.^[3] Oxidative stress is a harmful condition that occurs when there is an excess of ROS and a decrease in antioxidant levels, this may cause tissue damage by physical, chemical, psychological factors that lead to tissue injury in human and causes different diseases.^[4]

Antioxidant means "Against oxidation," antioxidants work to protect lipids from peroxidation by free radicals. Antioxidants are effective because, they are willing to give up their own electrons to free radicals.^[5] Antioxidants acts as a defense mechanism that protects against oxidative damage and include compounds to remove or repair damaged molecules. Medicinal plants are used worldwide in management of healthcare problems since time immemorial and approximately 60-80% of the world's population still depending on the

traditional medicine. Currently, the global demand of herbal medicine is increasing rapidly because of their safety margin and low cost. With the help of medicinal plants still many of the disease are getting cured.

Madhuca Indica commonly known as *Madhuca* belongs to the family of *Sapotaceae*, is an important economic plant growing throughout the subtropical region of the Indo-Pakistan subcontinent. Phytochemicals like carbohydrates, flavonoids, alkaloids, proteins, lipids, anthraquinones, coumarins, quinines, glycosides, terpenoids, cardiac glycosides, steroids and tannins were reported in this plant. Various parts of this plant are used as stimulants, demulcents, emollients, heating and astringents. The bark is a good remedy for itching, swellings, fractures and snake bites, as well as for diabetes mellitus. Mahua oil is used for the treatment of skin diseases, rheumatism, and headache and as a laxative. Fruits are astringent and largely employed as a lotion for chronic ulcers, in acute and chronic tonsillitis, and in pharyngitis.^[6,7]

Bark and leaves of *Madhuca indica* have reported to have antioxidant^[8,12], anticancer^[13], antimicrobial^[9,12], cytotoxic^[12], anti-inflammatory^[14], antipyretic.^[15] antidiabetic and antihyperglycemic^[16,18,11], anti-epileptic^[19], antinociceptive and antidiarrhoeal activities.^[20] Antiulcer activity^[21,22], hepatoprotective^[23] activity. In the present study, three different parts such as

wood, bark and leaves of *Madhuca indica* were selected to evaluate their antioxidant efficacy.

MATERIALS AND METHODS

Collection of plant materials

The plant materials used in the present study are wood, bark, leaf of *Madhuca indica*. The plants were collected from in and around Madhukkur, Thanjavur district, Tamil Nadu, India. The collected samples were carefully kept in polythene bags. They were dried in shade and stored in air tight containers until further studies.

Preparation of extracts

The dried wood, bark, leaf are powered using grinder, 60 gm of each powered was packed evenly in the soxhlet extractor and subjected to extraction with 210ml ethanol. After extraction, the solvent was distilled off and the extracts were concentrated on water bath to a dry residue and kept in a desiccator. The crude extract was used for phytochemical screening^[24] and *in vitro* antioxidant activity using various models such as Total antioxidant capacity^[25], DPPH scavenging activity^[26], Hydrogen peroxide scavenging activity^[27] and reducing power assay.^[28]

RESULTS AND DISCUSSION

Plants have been the basis of traditional medicines throughout the world for thousands of years and continue to provide new remedies to humankind; a great deal of effort has therefore focused on using available experimental to identify natural antioxidants from plants.^[29] In the proposed study, selected plant parts of wood, bark and leaf of *Madhuca indica* were collected and ethanol extracts were prepared and subjected to phytochemical screening and *in vitro* antioxidant activity. The obtained results were recorded and tabulated and discussed in this chapter.

Preliminary phytochemical screening

Plants constitute a source of novel chemical compounds which are of potential use in medicine and other applications. Plants contain many active compounds such as alkaloids, steroids, tannins, glycosides, volatile oils, fixed oils, resins, phenols and flavonoids. These active constituents are deposited in the specific parts of the plants such as leaves, flowers, bark, seeds, fruits, root, etc. These phytochemicals have much pharmacological potential including antioxidant, antibacterial^[30], antifungal^[31], antidiabetic^[32], anti-inflammatory^[33] and radio-protective activity^[34] and due to these properties those medicinal plants are largely used for medicinal purpose.

In the present study, qualitative analysis of different parts of *Madhuca indica* was investigated and represented in table 1. Carbohydrate, coumarins, cardiac glycosides, flavonoids, glycosides, quinones, steroids, tannins were present in wood extract. In bark extract, phytoconstituents of anthraquinones, alkaloids, cardiac glycosides, glycosides, quinones, steroids, phlobatannins and

terpenoids were identified. Leaf extract showed the presence of phytoconstituents of carbohydrates, flavonoids, tannins. Results revealed that ethanol wood extract of plant showed high amount of phytochemicals (8/15) followed by bark (7/15) and leaf extracts (3/15).

In vitro antioxidant activity

Under normal conditions, Reactive Oxygen Species (ROS) are continuously produced and converted to non reactive species by scavenging mechanism of different intracellular enzymatic and non-enzymatic antioxidant system.^[35] It is possible to reduce the risks of chronic diseases and prevent disease progression by either enhancing the body's natural antioxidant defenses or by supplementing with proven dietary antioxidants.^[36] Thus, a practical way to control these diseases is to increase the dietary intake of fruits and vegetables, which are rich sources of antioxidants.^[37,38] Since, the plant kingdom offers a wide range of natural antioxidants; recently there has been considerable interest in finding natural antioxidants from plant materials to replace synthetic ones. Moreover, these natural antioxidant substances are accepted to be secure and sound from the time when they occur in plant foods, and more enviable than synthetic one.

Hence, in the current study, wood, bark and leaf extract of *Madhuca indica* were subjected to analyze their antioxidant activity by various *in vitro* model such as total antioxidant activity, DPPH free radical scavenging assay, hydrogen peroxide scavenging activity and reducing power ability. Five different concentrations of (50, 100, 150, 200 and 250 µg/ml) wood, bark and leaf were evaluated for antioxidant activity. Antioxidant assay was also performed for standard ascorbic acid in order to compare the efficacy of selected plant parts. All the experiments were performed thrice and the results were averaged.

Total antioxidant activity

The total antioxidant capacity of the extracts were calculated based on the formation of the phosphomolybdenum complex which was spectrophotometrically measured at 695 nm. The total antioxidant activity of different concentrations (50, 100, 150, 200, 250 µg) of wood, bark and leaf extract of *Madhuca indica* were evaluated. Dose dependent antioxidant capacity was observed in all the selected parts and the percentage of antioxidant activity was found to be the range of 15.5% to 38.9%, 21.6% to 41.6% and 19.9% to 40.9% for wood, bark and leaf respectively. Percentage of antioxidant activity of ascorbic acid was found to be the range of 21.6% to 46.2% (table 2). Among the three different parts, bark extract exhibited maximum antioxidant activity but it was less than that of standard ascorbic acid. IC₅₀ values were found to be 135µg/ml, 160µg/ml, 150 µg/ml and 140µg/ml for wood, bark, leaves and ascorbic acid respectively.

DPPH free radical scavenging assay

DPPH a stable free radical and any molecule that can donate an electron or hydrogen to DPPH, can react with it and thereby bleach the DPPH absorption. DPPH is a purple colour dye having absorption maxima of 517 nm and upon reaction with a hydrogen donor the purple colour fades or disappears due to conversion of it to 2, 2-diphenyl-1-picryl hydrazine resulting in decrease in absorbance.

In our study, wood, bark and leaf extracts showed the radical scavenging activity of 42.2%, 51.2% and 44.5% respectively at 250 µg/ml, whereas at the same concentration, ascorbic acid exhibited 46.2% inhibition (table 4). Bark extract exhibited highest radical scavenging activity followed by standard ascorbic acid, leaf and wood. IC₅₀ of wood, bark and leaf extracts were found to be 140 µg/ml, 150µg/ml, 145µg/ml and 140µg/ml respectively which indicate the potency of scavenging activity of plant extracts. Standard ascorbic acid was found to have an IC₅₀ 140 µg/ml.^[39]

Hydrogen peroxide scavenging activity

Hydrogen peroxide itself is not particularly reactive with most biologically important molecules, but it is an intracellular precursor of hydroxyl radicals which is very toxic to the cell.^[40] In the present study, different concentrations of wood, bark and leaf of *Madhuca indica* were subjected to hydrogen peroxide scavenging activity. The result showed the dose dependent response for all the extract. Wood extract showed the inhibition range of 15.6 -38.9%, bark extract showed the inhibition range of 21.6 - 41.6% and for the leaf extract the inhibition

percentage were ranging from 19.9 to 40.9. Percentage of inhibition was found to be 21.6 - 46.2% for standard ascorbic acid (table 5).

IC₅₀ value of wood, bark and leaf were 135µg/ml, 160µg/ml, 150µg/ml respectively. Standard showed the IC₅₀ value 140µg/ml. Among the parts, bark showed the highest radical scavenging activity followed by leaf and wood but it was slightly less than that of standard ascorbic acid at 250 µg/ml. Plant extracts scavenged hydrogen peroxide, which might be attributed to the presence of phenolic group that donate electrons to hydrogen peroxide there by neutralizing it into water.^[41]

Reducing power assay

Reducing power of the fractions was assessed using ferric to ferrous reducing activity as determined spectro photometrically from the formation of Perl's Prussian blue color complex.^[42] Reducing power of different parts of *Madhuca indica* was evaluated in the current study which was compared with ascorbic acid. Highest reducing power of 0.25%, 0.51% and 0.42% were observed for wood, bark, leaf and for standard it was 46.2% at 250 µg/ml. The results confirmed the dose dependent response of all the parts of *Madhuca indica*. Among the parts, bark extract exhibited the potent reducing power. Moderate activity was observed in leaf extract and least activity was showed in wood extract of *Madhuca indica*. Plant extracts exhibited minimum activity than that of standard. IC₅₀ value of wood, bark and leaf were 25µg/ml, 20µg/ml and 40µg/ml respectively. Standard showed the IC₅₀ values 140µg/ml.

Table 1: Preliminary phytochemical screening of *Madhuca indica*.

S.No.	Phytochemicals	Results		
		Wood	Bark	Leaf
1	Anthraquinones	-	+	-
2	Alkaloids	-	+	-
3	Carbohydrates	+	-	+
4	Coumarins	+	-	-
5	Cardiac glycosides	+	+	-
6	Flavonoids	+	-	+
7	Glycosides	+	+	-
8	Phlobatannins	-	+	-
9	Phenols	-	-	-
10	Quinones	+	+	-
11	Saponins	-	-	-
12	Steroids	+	+	-
13	Triterpenoids	-	-	-
14	Terpenoids	-	+	-
15	Tannins	+	-	+

+ indicates presence

- indicates absence

Table 2: Total antioxidant activity of *Madhuca indica*.

S. No.	Concentration (µg/ml)	Total antioxidant activity in %			
		Plant extracts			Standard Ascorbic acid
		Wood	Bark	Leaf	
1	50	15.6±0.8	21.6±2.5	19.9±0.5	21.6±0.8
2	100	22.2±0.5	25.5±0.9	22.2±0.5	30.5±0.3
3	150	30.3±1.05	29.8±0.7	30.3±1.0	35.7±1.2
4	200	34.16±1.5	35.6±0.7	35.1±1.5	40.4±2.3
5	250	38.9±1.7	41.6±1.7	40.9±1.7	46.2±0.1
IC₅₀ value (µg/ml)		135	160	150	140

All values are expressed as mean ±S.D for three determinations.

Table 3: DPPH scavenging activity of *Madhuca indica*.

S.No.	Concentration (µg/ml)	Reducing power assay in %			
		Plant extracts			Standard Ascorbic acid
		Wood	Bark	Leaf	
1	50	0.11±0.007	0.23±0.02	0.12±0.007	21.6±0.8
2	100	0.14±0.10	0.27±0.007	0.22±0.007	30.5±0.3
3	150	0.18±0.007	0.33±0.02	0.28±0.007	35.7±1.2
4	200	0.22±0.03	0.43±0.06	0.37±0.002	40.4±2.3
5	250	0.25±0.01	0.51±0.007	0.42±0.004	46.2±0.1
IC₅₀ value (µg/ml)		25	20	40	140

All values are expressed as mean ±S.D for three determinations.

Table 4: Hydrogen peroxide scavenging activity of *Madhuca indica*.

S.No.	Concentration (µg/ml)	DPPH scavenging activity in %			
		Plant extracts			Standard Ascorbic acid
		Wood	Bark	Leaf	
1	50	16.9±0.51	18.60±2.8	19.3±5.1	21.6±0.8
2	100	23.2±1.2	26.20±3.1	24.8±2.4	30.5±0.3
3	150	31.3±1.2	34.03±1.45	31.1±1.3	35.7±1.2
4	200	33.±1.10	40.10±4.9	37.5±3.6	40.4±2.3
5	250	42.20±0.6	51.20±4.8	44.5±0.92	46.2±0.1
IC₅₀ value (µg/ml)		140	150	145	140

All values are expressed as mean ±S.D for three determinations.

Table 5: Reducing power assay of ethanol extract of *Madhuca indica*.

S.No.	Concentration (µg/ml)	Hydrogen peroxide scavenging activity in %			
		Plant extracts			Standard Ascorbic acid
		Wood	Bark	Leaf	
1	50	15.6±0.8	21.6±2.5	19.9±0.5	21.6±0.8
2	100	22.2±0.5	25.5±0.9	22.2±0.5	30.5±0.3
3	150	30.3±1.05	29.8±0.7	30.3±1.0	35.7±1.2
4	200	34.16±1.5	35.6±0.7	35.1±1.5	40.4±2.3
5	250	38.9±1.7	41.6±1.7	40.9±1.7	46.2±0.1
IC₅₀ value (µg/ml)		135	160	150	140

All values are expressed as mean ±S.D for three determinations.

CONCLUSION

From the result, it can be concluded that bark extract of *Madhuca indica* showed the highest antioxidant activity than leaf and wood extracts due to the presence of high number of phytochemicals. Further studies are needed to elucidate the *in vivo* antioxidant potential of the bark extract of *Madhuca indica* in the treatment of free radical induced human disease.

REFERENCES

- Gilbert D L. Fifty years of radical ideas, Ann Ny Acad Sci, 2000; 14: 899- 00.
- Kolayli S, Kucuk M, Yaylachi F, Dural N, and Vsta M, Comparison of *in vitro* antioxidant properties of trunk, bark of some tree species, proceedings of ICNP, 2002; 4(2): 133-139.
- Nakayama T, Yamada M, Osawa T, and Kawakishi S, Suppression of active oxygen induced

- cytotoxicity by flavonoids, *Bio Chem Pharmacol*, 1998; 45: 265-26.
- Valko M, Leibfritz D, Moncol J, Cronin MTD, Mazur M, Telser J, Free radicals and antioxidants in normal physiological functions and human disease, *International Journal of Biochemistry and Cell Biology*, 2007; 39(1): 44-84.
 - Kaczmarek M, Wojcicki J, Samochowiec L, Dutkiewicz T, and Sych Z, In Pharmacology: The influence of exogenous antioxidants and physical exercise on some parameters associated with production and removal of free radicals, *Journal of Medicinal Plants Research*, 1993; 54: 303-306.
 - Kirtikar KR, and Basu B.D, Indian medicinal plants. Sudhindra Nath Basu, Allahabad India, 1995; 747.
 - Khaleque A, Wahed MMA, Huq M.S, and Khan NA, *Madhuca latifolia*. I. Constituents of the seeds, *Science Research (Dacca, Pakistan)*, 1969; 6: 227-228.
 - Prashanth S, Kumar AA, Madhub B, and Yennamaneni PK, Antihyperglycemic and antioxidant activity of ethanolic extract of *Madhuca longifolia* bark, *Int J Pharma Sci Rev Res*, 2010; 5: 89-94.
 - Kaushik P, Kaushik D, Khokra SI, Sharma C, Aneja KR, Khah S, Mehra R, and Singh G, Evaluation of antioxidant and antimicrobial activity of *Madhuca indica*, *Pharmacology online*, 2012; 2: 1-8.
 - Patil AP, Patil VV, and Patil VR, *In vitro* free radicals scavenging activity of *Madhuca indica* Gmel, *Pharmacology online*, 2009; 2: 1344-1352.
 - Kumar A, Kumar A, Kumar S, Chauhan PK, Patil S, and Sharma P, *In vitro* evaluation of Phytochemical, antioxidant and antimicrobial activity of *Madhuca indica* leaf extract, *Asian Journal of Biochemical and Pharmaceutical Research*, 2011; 4(1): 175-185.
 - Bulbul J., and Begum Y., Antibacterial, Cytotoxic and Antioxidant activities of *Madhuca indica*, *Scientific Research Journal (SCIRJ)*, 2014; 2(4): 15-20.
 - Sangameswaran B, Saluja MS, and Sharma A, Anticancer activity of ethanol extract of *Madhuca longifolia* against Ehrlich Ascites Carcinoma, *Mol Clini Pharmacol*, 2012; 2: 12-19.
 - Vijayabhaskar K, and Chaitanyaprasad K, Anti-inflammatory and analgesic activities on leaves methanolic extract of *Madhuca indica* linn in wistar albino rats, *IJPCBS*, 2014; 4(2): 268-271.
 - Neha S, and Rekha V, Investigation of anti-inflammatory, analgesic and antipyretic properties of *Madhuca indica* Gmel, *Int J Mol Med Adv Sci*, 2010; 6: 26-30.
 - Akash P, Dahake S, Chirantan Rita C, and Prashant B., Antihyperglycemic activity of methanolic extract of *Madhuca longifolia* bark, *Scientific Research Journal*, 2010; 39: 3-8.
 - Samaresh PR, Shirole D, Tushar P, Shastry CS, Gheewala N, Goutam S, Ramachandra S, and Rajendra SV, Antioxidant and hepatoprotective activity of *Madhuca longifolia* (Koenig) bark against CCl_4 - induced hepatic injury in rats: *In vitro* and *In vivo* studies, *Res J Pharmaceut Biol Chem Sci.*, 2010; 1: 1-10.
 - Ghosh R, Dhande I, Kakade VM, Vohra RR, Kadam VJ, and Mehra, Antihyperglycemic activity of *Madhuca longifolia* in alloxan-induced diabetic rats, *Int J Pharmacol*, 2009; 6: 1-12.
 - Sandip P, Patel S, and Patel V, Investigation into the Mechanism of action of *Madhuca longifolia* for its anti-epileptic activity, *Pharmacognosy Communications*, 2011; 1: 18-22.
 - Rahman MA, Haque ME, Solaiman M, and Saifuzzaman M, Antinociceptive and antidiarrhoeal activities of *Madhuca indica* j. F. Gmel, *Pharmacology online*, 2011; 1: 473-480.
 - Mohod SM, and Bodhankar SL, Evaluation of antiulcer activity of methanolic extract of leaves of *Madhuca indica* J. F. Gmel in Rats, *Pharmacology online*, 2011; 3: 203-213.
 - Patel N, Kumar P, Rai S, Singh MP, Pandey R, Shukla SS, Saraf S, Patel S, and Patel D, Gastric ulcer protective effect of *Madhuca indica* roxb bark in wistar rats, *IJPSR*, 2014; 5(9): 4051-4055.
 - Chaudhary A, Bhandari A, and Pandurangan A, Hepatoprotective activity of methanolic extract of *Madhuca indica* on carbon tetrachloride-induced hepatotoxicity in rats, *Pharmacology online*, 2011; 1: 873-880.
 - Harborne JB, *Phytochemical Methods*. Chapman and Hall: London, 1998.
 - Shirwakar A, Shirwakar AR, Rajendran K, Punitha IRS, *In Vitro* Antioxidant Studies on the Benzyl Tetra Isoquinoline Alkaloid Berberine, *Biol. Pharm. Bull.*, 2006; 29(9): 1906-1910.
 - Blois MS, Antioxidant determinations by the use of a stable free radical, *Nature*, 1958; 181: 1199-1200.
 - Tanwar A, Trivedi VB, Kazmi SM, and Kazmi SN, Comparative bactericidal activity of two angiosperms, *Bulletin of Botanical Society, University of Sagar*, 2011; 27-36.
 - Oyaizu M, Studies on products of browning reactions: antioxidative activities of products of browning reaction prepared from glucosamine, *Japanese Journal of Nutrition*, 1986; 44: 307-315.
 - Krishnaiah D, Sarbatly R, Nithyanandam R, A review of the antioxidant potential of medicinal plant species, *Food Bioprod. Process*, 2011; 89(3): 217-233.
 - Nair R, Kalariya T, Sumitra C, Antibacterial activity of some selected Indian medicinal flora, *Turkey Journal of Biology*, 2005; 29: 41-47.
 - Khan M, Wassilew SW, *Natural pesticides from the Neem tree and other tropical plants*. (Eds) Schmitterer H and Asher KRS, German, Digital verlag GmbH, 1987; 645-650.
 - Singh N, and Gupta M, Effect of ethanolic extract of *Syzygium cumini* seed powder on pancreatic islets of alloxan diabetic rats, *Indian Journal of Experimental Biology*, 2007; 45: 861-867.

33. Kumar A, Ilavarasan R, Jayachandran T, Deecaraman M, Kumar M.R, Aravindan P, Padmanabhan N, Krishan MRV, Anti inflammatory activity of *Syzygium cumini* seed, African Journal of Biotechnology, 2008b; 7(8): 941-943.
34. Jagetia GC, Baliga MS, and Venkatesh P, Influence of seed extract *Syzygium cumini* (Jamun) on mice exposed to different doses of γ -radiation, *J Radiat Res.*, 2005; 46(1): 59-65.
35. Shao HB, Chu LY, Lu ZH, and Kang CM, Primary antioxidant free radical scavenging redox signaling pathways in higher plant cells, *Int J Biol Sci*, 2008; 4: 8-14.
36. Stanner SA, Hughes J, Kelly CN, Review of the epidemiological evidence for the antioxidant hypothesis, *Public Health. Nutrition*, 2000; 7: 401-422.
37. Demo A, Petrakis C, Kefalas P, and Boskou D, Nutrient antioxidants in some herbs and Mediterranean plant leaves, *Food Res Int. J Agric Food Chem.*, 1998; 31: 351-354.
38. Proteggente AR, Pannala SA, Paganga G, Buren LV, Wagner E, and Wiseman S, The antioxidant activity of regularly consumed fruits and vegetables reflects their phenolic and vitamin C composition, *Free Radic Res*, 2002; 36: 217-233.
39. Havesteen B, Flavanoids, a class of natural products of high pharmacological potency, *Biochem. Pharmacol*, 1983; 30: 1141-1148.
40. Yen GC, Duh PD, Scavenging Effect of Methanolic Extracts of Peanut Hulls on Free Radical and Active Oxygen Species, *J. Agric. Food Chem.*, 1994; 42: 629-632.
41. Miyake T, and Shibamoto T, Antioxidative Activities of Natural Compounds Found in Plants, *J. Agric. Food Chem.*, 1997; 45(5): 1819-1822.
42. Yildirim A, Mavi A, Kara AA, Determination of antioxidant and antimicrobial activities of *Rumex crispus* L. extracts, *J. Agric Food Chem*, 2001; 49: 4083-4089.